Occurrence, Morpho-Histopathological Characterization, and Infection Dynamics of *Posthodiplostomum* sp. (Strigeidida: Diplostomidae) in Cyprinid Fish of the Ganga River

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2021. Occurrence, morpho-histopathological	N., BERA A.K., PRASAD K.P., SURESH V.R., DAS B.K. l characterization, and infection dynamics of midae) in cyprinid fish of the Ganga River. Folia
cyprinid fish, <i>Labeo catla</i> and <i>Pethia conchon</i> , morphometry and characteristic histopathologi encysted in musculocutaneous tissues, and well er and 1.02±0.02 by 0.75±0.02 mm in <i>L. catla</i> an multifocal hyperpigmented areas in the muscul focal necrosis, and an infiltration of mononuc examined, only 3 <i>L. catla</i> were found to be in 13 <i>P. conchonius</i> were found to be infected at the F prevalence of <i>Posthodiplostomum</i> sp. in <i>L. catl</i> infection rate. This is the first report of <i>Posthodip</i>	a sp. (Strigeidida: Diplostomidae) was reported in the <i>ius</i> and was identified based on clinical signs, cyst cal lesions. The parasite was oval-round in shape, neapsulated. The cyst was 1.02 ± 0.02 by 0.79 ± 0.02 mm d <i>P. conchonius</i> , respectively. Microscopically, the ocutaneous tissues showed pericystic melanization, clear leukocytes. Out of the 5,820 freshwater fish infected in October at the Balagarh and in August, 'arakka stretch of the Ganga River. The spatio-temporal <i>la</i> and <i>P. conchonius</i> was <1%, indicating a lower <i>clostomum</i> sp. infection in <i>L. catla</i> in the Ganga
Key words: Labeo catla, Pethia conchonius, bla	ick spot disease, Ganga River.
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	 BAITHA R., MANNA S.K., KOUSHLESH S.K., DAS J 2021. Occurrence, morpho-histopathological <i>Posthodiplostomum</i> sp. (Strigeidida: Diploston Biologica (Kraków) 69: 113-120. A metacercarial infection of <i>Posthodiplostomum</i> cyprinid fish, <i>Labeo catla</i> and <i>Pethia conchon</i> morphometry and characteristic histopathologi encysted in musculocutaneous tissues, and well er and 1.02±0.02 by 0.75±0.02 mm in <i>L. catla</i> an multifocal hyperpigmented areas in the muscul focal necrosis, and an infiltration of mononuc examined, only 3 <i>L. catla</i> were found to be in 13 <i>P. conchonius</i> were found to be infected at the F prevalence of <i>Posthodiplostomum</i> sp. in <i>L. catla</i> infection rate. This is the first report of <i>Posthodip</i> Ganga River. It is also the first report of <i>Posth</i> River. Key words: <i>Labeo catla</i>, <i>Pethia conchonius</i>, bla <i>Raju BAITHA</i>, <i>Sanjib Kumar MANNA</i>[™], <i>Satish K.</i> <i>Basanta Kumar DAS</i>, <i>ICAR-Central Inland Fishe</i> <i>Bengal, India.</i> <i>E-mail: sanjibmanna@yahoo.com; sanjib.manr.</i> <i>Kurcheti Pani PRASAD</i>, <i>ICAR-Central Institute op</i>

Young and juvenile cyprinid fish species constitute the largest susceptible group to being the second intermediate host of *Posthodiplostomum* (JALALI 1998; ROLBIECKI 2004; ONDRAČKOVÁ *et al.* 2004a; RAKAUSKAS & BLAŽEVIČIUS 2009; KARIMIAN *et al.* 2013). Clinically, the infection is identified by the appearance of black spots in different organs such as the fins, skin, muscles, gills, operculum, and buccal cavity of affected fish. Infected fish also show signs of emaciation, spinal deformation, and degenerative changes in musculature, hepatic and renal dystrophy, anorexia, retarded growth, and pathological changes in the blood and even succumb to death in severe cases (ROLBIECKI 2004). Histologically, pinheadsized black spots in the affected tissues show perifocal fibrous nodules bounded by a ring of melanin and an infiltration of inflammatory cells (TEIXEIRA-DE MELLO & EGUREN 2008; FERGUSON *et al.* 2010; FLORES-LOPES & THOMAZ 2011; NEGREA *et al.* 2015).

Freshwater digeneans often show seasonality in their infection pattern, which is related to cercarial emergence when water temperature increases (CHUBB 1979). The peak cercarial release coincides with the

© Institute of Systematics and Evolution of Animals, PAS, Kraków, 2021 Open Access article distributed under the terms of the Creative Commons Attribution License (CC-BY) <u>http://creativecommons.org/licences/by/4.0</u> breeding period of cyprinid fish therefore ensuring a chance for maximum infection in fish and hence parasite distribution by fish dispersal, as well as the maintenance of high parasite abundance (VLADIMIROV 1960). Further, metacercarial infection of juvenile fish provides the best chance of parasite transmission to the final piscivorous host (FISCHER & KELSO 1988). The infection pattern of Posthodiplostomum cuticola varies by fish species and season (ONDRAČKOVÁ et al. 2004a, b; IADRENKINA 2014; NEGREA et al. 2015), location on musculocutaneous tissue (NEGREA et al. 2015), type and zones of a water body, and their physico-chemical parameters (ONDRAČKOVÁ et al. 2004a; IADRENKINA 2014). The spatial, seasonal, and age-related variations in the prevalence of the diplostomid digenean, Posthodiplostomum-Ornithodiplostomum clade in valenciid fish have been reported (KALOGIANNI et al. 2017).

The Ganga River is the largest river in India and the fifth largest in the world. It flows through multiple agro-climatic zones and is home to 143 fish species with cyprinid fish constituting the major catch (53.47%) (SARKAR *et al.* 2011). Reports on fish diseases in feral populations are sporadic and scanty compared to those in aquaculture systems. Therefore, metacercarial infection of *Posthodiplostomum* in the

fish of the Ganga River has been investigated to develop the status of infection in rivers.

Materials and Methods

Study area and fish samples

Regular sampling surveys were conducted at four stations, viz., Farakka (24°47′38.478′′N, 87°55′26.413′′E), Jangipur (24°27'58.16''N, 88°03'57.05''E), Rejinagar (23°50'10.64''N, 88°13'55.60''E), and Balagarh (23°07'44.05''N, 88°27'58.04''E) (Fig. 1) in the lower stretch of the Ganga River, West Bengal, India. Fish samples were caught using gill nets (mesh 22-120 mm), cast nets (12-14 mm), and indigenous traps set along the shallow (depth: 0.5-2 m) and vegetated areas of the river and were identified following the keys described by JAYARAM (1999). The total length (TL) and weight (W) of the fish were measured to the nearest 1 mm with a digital calliper and 0.01 g on an electronic scale, respectively. The sampled fish were examined carefully for the presence of black spots that deviated from the natural color pattern of the fish. Live fish were euthanized with excess doses of clove oil before examination. All of the visibly in-

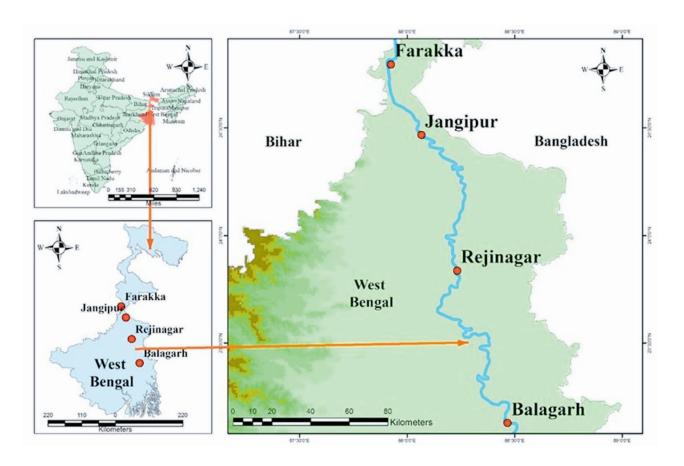


Fig. 1. Sampling sites along the lower stretch of the Ganga River.

fected and some of the non-infected fish samples were preserved in 10% neutral buffered formalin.

Parasitological examination

In the laboratory, infected fishes were examined thoroughly and the number of black spots in various body parts of individual fish was recorded. The encysted metacercariae were removed carefully using a pair of fine forceps from infected body parts, viz., the trunk muscles close to the head, the middle and caudal portions of a fish separately. Isolated cysts were washed vigorously in 0.85% physiological saline. Forty metacercarial cysts were picked randomly from the *Labeo catla* and *Pethia conchonius* specimens for morphometric characterization using a microscope equipped with a digital camera. Measurements (mm) of fresh cysts were taken with a calibrated eye-piece micrometer.

Histopathological analyses

The musculocutaneous tissues collected from infected fish were fixed in 10% neutral buffered formalin for histopathological examination. The fixed tissues were processed for histological examination using standard techniques, embedded in paraffin wax, sliced into 5 μ m thick sections, mounted, and then stained with haematoxylin and eosin following standard protocol (PRESNELL & SCHREIBMAN 1997).

The ecological aspects of infection dynamics, viz., mean prevalence, mean abundance, mean intensity, and range of infection were estimated following MARGOLIS *et al.* (1982) and BUSH *et al.* (1997). The standard statistical computation was carried out using Microsoft Excel (Office 2016).

Results and Discussion

Morphometric characterization of the parasite

Posthodiplostomum cuticola and Apophallus sp. (HOOLE et al. 2001), Uvulifer ambloplitis and Crassiphiala bulboglossa (WOO 2006) have been known to infect the early stages of a wide range of fish species and causing black spot disease. U. ambloplitis and C. bulboglossa are mainly known from temperate inland waters in the Northern hemisphere (PAPERNA & DZIKOWSKI 2006), Apophallus brevis is limited to the continent of North America, infecting only Yellow perch, Perca flavescens (SINCLAIR 1972), and P. cuticola is globally distributed (DÖNGES 1964; ONDRAČKOVÁ et al. 2004b; ZRNČIĆ et al. 2009; NGUYEN et al. 2012; KARIMIAN et al. 2013; ATHOKPAM & TANDON 2014).

Five thousand eight hundred and twenty (5,820) fish samples belonging to 30 families (Cyprinidae, Channidae, Schilbeidae, Bagridae, Ambassidae, Osphronemidae, Cobitidae, Heteropneustidae, Anabantidae, Gobiidae, Mastacembelidae, Sciaenidae, Engraulidae, Tetraodontidae, Siluridae, Clariidae, Belonidae, Mugilidae, Sisoridae, Nandidae, Notopteridae, Pangasidae, Synbranchidae, Chacidae, Badidae, Cynoglossidae, Soleidae, Clupeidae, Ailiidae, and Anguilidae) were examined from the studied stations in the lower stretch of the Ganga River. Among the several genera and species of the fish studied from the river, only two cyprinid species, namely L. catla and *P. conchonius*, were found to be parasitized by metacercariae of *Posthodiplostomum* sp. (Fig. 2a, 2b). Out of nearly six thousand fish samples examined, only sixteen (16) fish were found to be infected, in which the number of cysts varied widely, from 1-87 cysts, in each infected fish. The infected L. catla samples

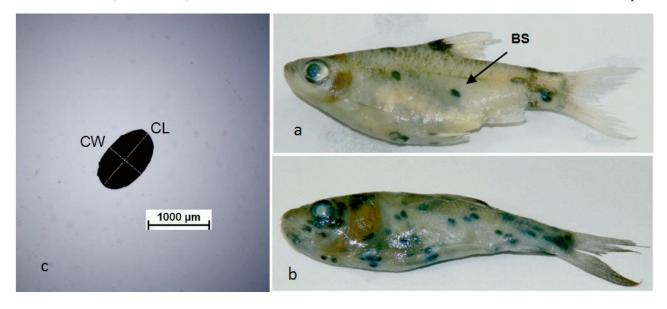


Fig. 2. Black spot disease in cyprinid fish from the lower stretch of the Ganga River: a - P. *conchonius*, b - L. *catla*., c - cyst, BS – black spot; CL - cyst length; CW - cyst width.

were juveniles with a mean length of 56.66 mm and mean weight of 2.52 g, while the mean length and weight of infected *P. conchonius* samples was 36.23 mm and 0.82 g, respectively.

In the present study, the metacercarial cysts were oval-round in shape (Fig. 2c) and 0.82-1.13 (1.02±0.02) by 0.63-1.05 (0.79±0.02) mm and 0.87-1.16 (1.02±0.02) by 0.6-0.91 (0.75±0.02) mm for *L. catla* and *P. conchonius*, respectively. For differential diagnosis, the morphometric comparison of the metacercariae of major Posthodiplostomum spp. that infect freshwater fish are presented in Table 1. A comparative description of the cyst dimensions of *Posthodiplostomum* sp. with morphologically similar parasitic species responsible for black spot disease in freshwater fish have also been tabulated (Table 2). The shape and dimensions of the cysts in the present investigation clearly indicated that the causative agent of black spot disease in P. conchonius and L. catla belonged to the genus Posthodiplostomum with a close resemblance to P. cuticola. Variations in the dimension of cysts may be correlated with the stage of development of the encysted digenetic trematodes as evidenced in case of C. bulboglossa (HOFFMAN 1956). However, similar studies on other digenetic trematodes are lacking.

Histopathologic characterization of the parasite

The metacercariae-infected musculocutaneous tissues of *L. catla* and *P. conchonius* showed a proliferation of melanocytes just beneath the epidermis and a melanization of the fibrous capsule surrounding the metacercariae (Figs 3a-3c) which were visible macroscopically as multifocal hyperpigmented areas (Fig. 2a) along with slightly raised nodular structures, seen in the case of L. catla (Fig. 2b). The surrounding muscle tissue suffered from medium to high levels of pressure atrophy, and marked degeneration as well as focal necrosis (Figs 3a-3c). An infiltration of numerous immune cells (mononuclear leukocytes) (Fig. 3d) could be observed in the fibrous capsule encircling the parasite. A melanization in the fibrous capsule surrounding the parasite could be seen (Figs 3a-3c). Because of the thick capsule, the thick-walled metacercariae of digenetic trematodes are able to evade host immune responses (GRACZYK 1997). The metacercarial cyst of A. brevis also has a bony component (SINCLAIR 1972) and a distinctive spiny tegument unique to Apophallus spp. (KENT et al. 2004). Further, encysted metacercariae of Apophallus sp. (Heterophyidae Leiper, 1909) can be distinguished from other black spot forming digenetic trematodes by their cyst bi-layer (a thin inner layer of parasitic origin and thick outer layer of host origin), fibrous outer layer, and occasional calcification (GIBSON 1996). This substantiates that the histological changes observed in the present study were not due to the metacercariae of heterophyid digenetic trematodes, mainly becaue the melanized capsule around the cyst was thinner. Crassiphiala bulboglossa (Crassiphialinae Sudarikov, 1960) has a loosely fitted, spacious and thinner cyst wall, mainly infects skin, and is occasionally found between myotomes and on gill arches (GIBSON 1996). The present study has recorded the infection in musculocutaneous tissue, but not on the gill arches of P. conchonius and L. catla. Metacercariae of Uvulifer Yamaguti, 1934 are fitted relatively

Table 1

Morphometric comparison of *Posthodiplostomum* spp. metacercarial cyst dimensions from freshwater fish

Host species	Parasitic species	Site of infection	Shape	Cyst dimensions (mm)	Reference
L. catla	Posthodiplostomum sp.	Muscle	oval-round	0.82-1.13 (1.02±0.02) by 0.63-1.05 (0.79±0.02)	Present study
P. conchonius	Posthodiplostomum sp.	Muscle	oval-round	0.87-1.16 (1.02±0.02) by 0.6-0.91 (0.75±0.02)	Present study
Channa punctata	Posthodiplostomum sp.	Muscle	oval-round	0.64-0.67 (0.65±0.01) by 0.53-0.55 (0.51±0.06)	ATHOKPAM & TANDON 2014
Channa argus	Posthodiplostomum sp.	Trunk muscle	oval-round	0.79-1.40 (1.04±0.18) by 0.54-0.94 (0.69±0.14)	NGUYEN et al. 2012
Ctenopharyngodon idella	P. cuticola	Skin, fins, oral cavity	round	0.900-0.990 (0.946±0.025)	MAJA et al. 2012
Aristichthys nobilis	P. cuticola	Skin, fins, oral cavity	round	0.900-0.980 (0.944±0.023)	MAJA et al. 2012
Carassius gibelio	P. cuticola	Skin, Musculature	spherical-oval	0.734-0.918 by 0.286-0.306	SUKHEROVA et al. 2005
Phoxinus laevis	P. cuticola	Muscle, skin, fins	oval-round	0.80-0.96 (0.89±0.05) by 0.69-0.88 (0.81±0.05)	DÖNGES 1964 (cited in NGUYEN et al. 2012)

Table 2

Comparative description of the metacercarial cyst dimensions of *Posthodiplostomum* species with morphologically similar parasitic species responsible for black spot disease in freshwater fish

Causative agent	Host/family	Site of infection	Cyst dimensions (mm)	Reference
Posthodiplostomum sp.	L. catla		0.82-1.13 (1.02±0.02) by 0.63-1.05 (0.79±0.02)	Present study
Posthodiplostomum sp.	P. conchonius	Muscle	0.87-1.16 (1.02±0.02) by 0.6-0.91 (0.75±0.02)	Present study
P. cuticola	Phoxinus laevis	Muscle, skin, fins	0.80-0.96 (0.89±0.05) by 0.69-0.88 (0.81±0.05)	Dönges 1964
P. cuticola	Cyprinidae	Skin	0.69-0.99 (Dia.)	HOOLE et al. 2001
Apophallus sp.	Cyprinidae	Skin	0.14-0.28 (Dia.)	HOOLE et al. 2001
U. ambloplitis	Pike, Minnows, Small-mouth bass, Bluegill, Sunfish, Western mosquitofish, and Bluegill sunfish	Muscle	0.377 by 0.2	Hickner 2011
C. bulboglossa	Ecosidae, Cyprinidae, Cypri- nodontidae, Etheostomidae, Percidae, and Umbridae	Muscle	0.31-0.4 by 0.18-0.215	Hoffman 1956

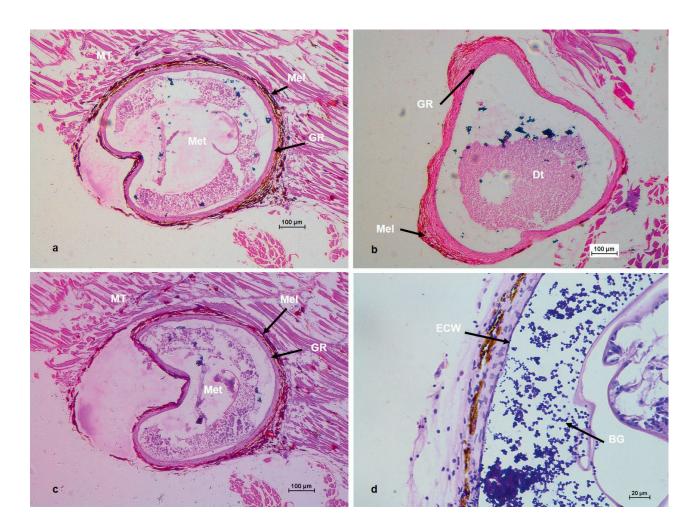


Fig. 3. Light micrographs of the metacercariae of *Posthodiplostomum* sp. in the musculocutaneous tissue: a-b – *P. conchonius* and c-d – *L. catla* fingerling, MT – musculocutaneous tissue, Mel – melanocytes, BG – basophilic granula, ECW – eosinophilic cyst wall; Met – metacercariae; GR – granulomatous reaction; Dt – degenerating metacercariae in distorted section. H&E staining.

tightly in cysts and the cysts are oval-pyriform and infect the fins, skin, and muscles of host fish. Thus, based on the location of the cysts on the body, the characteristics and histopathological lesions, the metacercariae infecting *P. conchonius* and *L. catla* could be *Posthodiplostomum* sp.

Infection pattern

Among fish species belonging to 30 families investigated in the present study, the occurrence of metacercarial *Posthodiplostomum* sp. was observed only in juveniles of *P. conchonius* and *L. catla*. The cysts of *Posthodiplostomum* sp. infected the epidermal layer of the musculocutaneous tissue. The present study is well supported by earlier observations that young and juvenile cyprinid fish species constitute the largest susceptible group to being the second intermediate host *of Posthodiplostomum* sp. (ROLBIECKI 2004; ONDRAČKOVÁ *et al.* 2004a; RAKAUSKAS & BLAŽEVIČIUS 2009; ZRNČIĆ *et al.* 2009; KARIMIAN *et al.* 2013).

The sampling period and station-wise prevalence, mean intensity, mean abundance, and range of infection are given in Table 3, and the same have been calculated at species level within family (Table 4). Table 3

Table 3

Sampling period and station-wise infection pattern of *Posthodiplostomum* sp. in the lower stretch of the Ganga River

Fish species	Items	Sampling periods						Sampling stations			
T ISH Species	Items	Aug-16	Oct-16	Dec-16	Feb-17	Apr-17	Jun-17	Farakka	Jangipur	Rejinagar	Balagarh
	Ν	1455	902	650	960	730	1123	2530	695	1150	1445
	n	_	3	_	-		_	_	_	_	3
	Q	_	214	_	_	_	_	_	_	_	214
L. catla	Р	_	0.33	_	_	-	_	_	_	_	0.2
	MI	_	71.33	_	_	_	_	_	_	_	71.33
	MA	_	0.23	_	_	_	_	_	_	_	0.14
	R	_	62-87	_	_	_	_	_	_	_	62-87
P. conchonius	N	1455	902	650	960	730	1123	2530	695	1150	1445
	n	13	_	_	_	_	_	13	_	_	_
	Q	67	—	_	_	_	_	67	_	_	_
	Р	0.89	_	_	_	_	_	0.51	_	_	_
	MI	5.15	_	_	_	_	_	5.15	_	_	_
	MA	0.04	_	_	_	_	_	0.02	_	_	_
	R	1-10	_	_	_	_	_	1-10	_	_	_

Number of fish examined at each station and sampling period (N), number of infected fish specimens (n), number of metacercariae recovered (Q), prevalence (P), mean intensity (MI), mean abundance (MA), and range of infection (R)

Table 4

Intra-familial infection pattern of Posthodiplostomum sp. in the lower stretch of the Ganga River

	Samples	Fish species								
Parasite		Labeo rohita	Labeo calbasu	Labeo catla	Cirrhinus mrigala	Puntius ticto	Puntius sarana	Puntius sophore	Pethia conchonius	
	Ν	93	170	66	55	759	285	1955	554	
Posthodiplostomum sp.	n	_	_	3	_	_	_	_	13	
	Q	_	_	214	_	_	_	_	67	
	Р	_	_	4.54	_	_	_	_	2.3	
	MI	_	_	71.33	_	_	_	_	5.15	
	MA	_	_	3.24	_	_	_	_	0.12	
	R	_	_	62-67	_	-	_	_	1-10	

Number of fish examined at each station and sampling period (N), number of infected fish specimens (n), number of metacercariae recovered (Q), prevalence (P), mean intensity (MI), mean abundance (MA), and range of infection (R).

shows that P. conchonius at Farakka and L. catla at Balagarh were infected with the metacercarial stage of Posthodiplostomum sp. in August and October, respectively. Table 4 shows that P. conchonius and L. catla were the only species infected with the metacercarial stage of Posthodiplostomum sp. from among various cyprinid fish species. Labeo catla was infected more heavily than P. conchonius. The overall and intra-familial level of prevalence, mean intensity, mean abundance, and range of infection were higher for L. catla than P. conchonius. The period of August and October is monsoon and post-monsoon season, respectively on the east coast of India, which are also the periods of fish breeding and high foraging activity of younger fishes in shallow, vegetated areas of overflown river. This period also coincides with the breeding of aquatic snails and the aggregation of piscivorous birds in the same locality. The presence of all the intermediate and final host species of Posthodiplostomum favor completion of its life cycle (ONDRAČKOVÁ et al. 2004; TEIXEIRA-DE MELLO & EGUREN 2008; ZRNČIĆ et al. 2009; FLORES-LOPES & THOMAZ 2011).

Conclusion

Posthodiplostomum sp. (Strigeidida: Diplostomidae) infection was identified based on the clinical signs, comparative cyst morphometry, and the characteristic histopathological changes in the fish host. This is the first report of infection by the metacercariae of *Posthodiplostomum* sp. (Strigeidida: Diplostomidae) in *P. conchonius* and in the Ganga River. It is also the first report of infection in *L. catla* in the Ganga River. However, a less than 1% prevalence of *Posthodiplostomum* sp. in *L. catla* and *P.conchonius*, indicates a low infection rate.

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Author Contributions

Research concept and design: R.B., S.K.M., A.K.B., K.P.P., S.V.R., B.K.D.; Collection and/or assembly

of data: R.B., S.K.K., N.D.; Data analysis and interpretation: R.B., S.K.M., S.K.K., N.D., A.K.B.; Writing the article: R.B.; Critical revision of the article: R.B., S.K.M., S.K.K., A.K.B., K.P.P., S.V.R., B.K.D.; Final approval of article: B.K.D.

Conflict of Interest

The authors declare no conflict of interest.

References

- ATHOKPAM V.D., TANDON V. 2014. Morphological and molecular characterization of *Posthodiplostomum* sp. (Digenea: Diplostomidae) metacercaria in the muscles of snakeheads (*Channa punctata*) from Manipur, India. Helminthol. **51**(2): 141-152. http://dx.doi.org/10.2478/s11687-014-0221-z
- BUSH A.O., LAFFERTY K.D., LOTZ J.M., SHOSTAK A.W. 1997. Parasitology meets ecology on its own terms: MARGOLIS *et al.* revisited. J. Parasitol. Res. 575-583. https://doi.org/10.2307/3284227
- CHUBB J.C. 1979. Seasonal occurrence of helminths in freshwater fishes part II. Trematoda. (In: Advances in parasitology, 17, W.H.R. LUMSDEN, R. MULLER, J.R. BAKER ed. Academic Press, London): 141-313.
- DÖNGES J. 1964. The life cycle of *Posthodiplostomum cuticola* (von Nordmann 1832) Dubois 1936 (Trematoda, Diplostomatidae). Zeitschrift für Parasitenkunde **24**(2): 169-248.
- FERGUSON J.A., SCHRECK C.B., CHITWOOD R., KENT M.L. 2010. Persistence of infection by metacercariae of *Apophallus* sp., *Neascus* sp., and *Nanophyetus salmincola* plus two myxozoans (*Myxobolus insidiosus* and *Myxobolus fryeri*) in coho salmon *Oncorhynchus kisutch*. J. Parasitol. **96**(2): 340-347. https://doi.org/10.1645/GE-2289.1
- FISCHER S.A., KELSO W.E. 1988. Potential parasite-induced mortality in age-0 bluegills in a floodplain pond of the lower Mississippi River. Trans. Am. Fish. Soc. **117**: 565-573.
- FLORES-LOPES F., THOMAZ A.T. 2011. Assessment of environmental quality through analysis of frequency of the black spot disease in an assemblage of fish, Guaíba Lake, RS, Brazil. Braz. J. Biol. **71**: 915-923. https://doi.org/10.1590/S1519-69842011000500012
- GRACZYK T.K. 1997. Immunobiology of trematodes in vertebrate hosts. Advances in trematode biology, 384-398.
- HICKNER M. 2011. Uvulifer ambloplitis (On-line), Animal Diversity Web. Accessed September 15, 2021 at https://animaldiversity.org/accounts/Uvulifer_ambloplitis/
- HOFFMAN G.L. 1956. The life cycle of *Crassiphiala bulboglossa* (Trematoda: Strigeida). Development of the metacercariae and cyst, and effect on the fish hosts. J. Parasitol. **42**: 435-444.
- HOOLE D., BUCKE D., BURGESS P., WELLBY I. 2001. Diseases of carp and other cyprinid fishes. Fishing news books, Blackwell Science Ltd, Oxford. Pp. 63-122.
- IADRENKINA E.N. 2014. Differences in the infestation rate of young cyprinid fishes (Cypriniformes) by metacercaria of *Posthodiplostomum cuticola* (Digenea, Diplostomatidae) in river and lake systems of the Lake Chany basin (Western Siberia). Parazitologiia **48**: 234-244.
- JALALI B. 1998. The parasites and parasitic diseases of freshwater fishes of Iran. Fisheries Co of Iran. 564.
- JAYARAM K.C. 1999. The freshwater fishes of Indian region. Narendra Publishing House, Delhi.
- KALOGIANNI E., KMENTOVÁ N., HARRIS E., ZIMMERMAN B., GIAKOUMI S., CHATZINIKOLAOU Y., VANHOVE M.P. 2017. Occurrence and effect of trematode metacercariae in two endan-

gered killifishes from Greece. Parasitol. Res. **116**(11): 3007-3018. https://doi.org/10.1007/s00436-017-5610-z

- KARIMIAN E., GHORBANI R., HAJIMORADLOU A. 2013. First occurrence and intensity of *Posthodiplostomum cuticola* (Nordmann 1832) (Digenea; Diplostomatidae) metacercariae in monkey goby (*Neogobius pallasi* Berg 1916) in the Zarringol stream, Golestan Province, Iran. Glob. Vet. **10**: 505-510. http://dx.doi.org/10.5829/idosi.gv.2013.10.5.20413
- KENT V.M.L., WATRAL V.G., WHIPPS C.M., CUNNINGHAM M.E., CRISCIONE C.D., HEIDEL J.R., CURTIS L.R., SPITSBERGEN J., MARKLE D.F. 2004. A digenean metacercaria (*Apophallus* sp.) and a myxozoan (*Myxobolus* sp.) associated with vertebral deformities in cyprinid fishes from the Willamette River, Oregon. J. Aquat. Anim. Health. 16: 116-129. https://doi.org/10.1577/H04-004.1
- GIBSON D.I. 1-996. Trematoda (In: Guide to the Parasites of Fishes of Canada. Part IV. L. MARGOLIS, Z. KABATA eds, NCR Research Press, Ottawa. Can. Spec. Publ. Fish. Aquat. Sci. **124**: 1-373.
- MARGOLIS L., ESCH G.W., HOLMES J.C., KURIS A.M., SCHAD G. 1982. The use of ecological terms in parasitology (report of an ad hoc committee of the American Society of Parasitologists). J. Parasitol. **68**(1): 131-3.
- NEGREA O., MIREŞAN V., RĂDUCU C., ONACIU G., NEGREA O., LAŢIU C., DANIEL C. 2015. Some investigations on incidence and infestation level in Cyprinid postodiplostomosis. Bulletin of University of Agricultural Sciences and Veterinary Medicine Cluj-Napoca. Animal Science and Biotechnologies, 72(2): 203-206. http://dx.doi.org/10.15835/buasvmcn-asb:11457
- NGUYEN T.C., LI Y.C., MAKOULOUTOU P., JIMENEZ L.A., SATO H. 2012. *Posthodiplostomum* sp. metacercariae in the trunk muscle of Northern snakeheads (*Channa argus*) from the Fushinogawa River, Yamaguchi, Japan. J. Vet. Med. Sci. **74**: 1367-1372. http://dx.doi.org/10.1292/jvms.12-0025
- ONDRAČKOVÁ M., REICHARD M., JURAJDA P., GELNAR M. 2004a. Seasonal dynamics of *Posthodiplostomum cuticola* (Digenea, Diplostomatidae) metacercariae and parasite-enhanced growth of juvenile host fish. Parasitol. Res. **93**: 131-136. https://doi.org/10.1007/s00436-004-1123-7
- ONDRAČKOVÁ M., ŠIMKOVÁ A., GELNAR M., JURAJDA P. 2004b. *Posthodiplostomum cuticola* (Digenea: Diplostomatidae) in intermediate fish hosts: factors contributing to the parasite infection and prey selection by the definitive bird host.

Parasitol. 129: 761-770.

https://doi.org/10.1017/s0031182004006456

- PAPERNA I., DZIKOWSKI R. 2006. Digenea (Phylum Platyhelminthes). (In: Fish diseases and disorders. P.T.K. WOO ed. CABI, Oxfordshire): 345-390.
- PRESNELL J.K., SCHREIBMAN M.P., HUMASON G.L. 1997. Humason's animal tissue techniques. Johns Hopkins University Press, Canada.
- ROLBIECKI L. 2004. Distribution of *Posthodiplostomum cuticola* (Nordmann, 1832) (Digenea; Diplostomidae) metacercariae in cyprinids of the Vistula lagoon, Poland. Arch. Pol. Fish. **12**: 93-98.
- SARKAR U.K., PATHAK A.K., SINHA R.K., SIVAKUMAR K., PANDIAN A.K., PANDEY A., DUVEY V.K., LAKRA W.S. 2011. Freshwater fish diversity in the River Ganga (India): changing pattern, threats and conservation perspectives. Rev. Fish Biol. Fish. 22: 251-272. http://dx.doi.org/10.1007/s11160-011-9218-6
- SINCLAIR N.R. 1972. Studies on the heterophyid trematode *Apophallus brevis*, the "sand-grain grub" of yellow perch (*Perca flavescens*). II. The metacercaria: position, structure, and composition of the cyst; hosts; geographical distribution and variation. Can. J. Zool. **50**: 577-584.
- TEIXEIRA-DE MELLO F., EGUREN G. 2008. Prevalence and intensity of black-spot disease in fish community from a subtropical stream (Santa Lucíariver basin, Uruguay). Limnetica, **27**: 251-258. https://doi.org/10.23818/limn.27.20
- VLADIMIROV V.L. 1960. The morphology and biology of the cercariae of *Posthodiplostomum cuticola* (Nordmann 1832) Dubois, 1936-causing black-spot disease in fish. Dokl. Akad. Nauk SSSR, **135**(4): 1009-1011.
- RAKAUSKAS V., BLAŽEVIČIUS Č. 2009. Distribution, prevalence and intensity of roach (*Rutilus rutilus* (Linnaeus, 1758)) parasites in inland waters of Lithuania in 2005-2008. Acta zool. litu. **19**: 99-108. https://doi.org/10.2478/v10043-009-0017-4
- WOO P.T.K. 2006. Fish diseases and disorders: 1. Protozoan and metazoan infections. 2nd eds. CAB International, Oxfordshire. Pp. 791.
- ZRNČIĆ S., ORAIĆ D., MIHALJEVIĆ Ž., ĆALETA M., ZANELLA D., JELIĆ D., JELIĆ M. 2009. First observation of *Posthodiplo-stomum cuticola* (Nordmann, 1832) metacercariae in cypriniformes from Croatia. Helminthologia, **46**: 112-116. https://doi.org/10.2478/s11687-009-0022-y