Short Note

The First Record of Nucleopolyhedrovirus Isolated from the Satin Moth Leucoma salicis L. (Lepidoptera, Lymantriidae) in Turkey

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A nucleopolyhedrovirus was isolated and characterized for the first time from Leucoma salicis L. (Lepidoptera, Lymantriidae) in Turkey. The virus was observed in populations of L. salicis in Gümüşhane. The dimensions of the polyhedrae fell between 2.08 ± 0.31 (1.51-2.64) μ m (N=50). Virions contain 2 to 15 nucleocapsids per virion as seen in cross-section of polyhedrae. The sizes of the viral particles ranged between 250-290 x 32-40 nm. The virus was determined as a Turkish isolate of Leucoma salicis nucleopolyhedrovirus (LesaNPV-TR).

Key words: Leucoma salicis L. (Lepidoptera, Lymantriidae), nucleopolyhedrovirus, Turkish isolate, biological control.

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The satin moth *Leucoma salicis* L. (Lepidoptera, Lymantriidae), previously known as Stilpnotia salicis, is a serious defoliator occurring throughout Europe and Asia (LIPA & ZIEMNICKA 1996). It is also an important pest in Turkey (ÇANAKÇİOĞLU 1983). Satin moth larvae feed on all species of poplar and willow (*Populus* spp.), but also on oak and crabapple (JAKUBOWSKA et al. 2005). Severe feeding damage results in reduced growth of stems and finally tree mortality (LANGOR 1995). Increasing problems with resistance of this pest to most commonly used synthetic insecticides has spurred the search for alternative pest management strategies that reduce reliance on synthetic insecticides. Fortunately, L. salicis populations are reduced by a natural nucleopolyhedrovirus (NPV) (Baculovirus) disease (ZIEMNICKA 1981). This virus is capable of causing disease in the larvae of L. salicis, but does not harm other animals or plants. Nucleopolyhedroviruses represent an effective, selective and safe biological control agent (GREATHEAD 1976). The best results with nucleopolyhedroviruses have been achieved against lepidopteran pests, for example L. salicis using LesaNPV (ZIEMNICKA 1981; 2000). Several important studies have been carried out on the Polish isolate of Leucoma salicis nucleopolyhedrovirus (STROKOVSKAYA et al. 1996;

LIPA & ZIEMNICKA 1996; JAKUBOWSKA et al. 2005). Some isolates of this virus were recorded in part of Europe (WEISER et al. 1954; SKATULLA 1985) but there is no record of the isolation and characterization of this virus from Asia. Asia is a potential source of new and interesting NPV strains (MURILLO et al. 2001). Some strains of nucleopolyhedrovirus isolated from different geographic localities may present better insecticidal activities, which make them more suitable for host control and show important differences in biological activity (MURILLO et al. 2001).

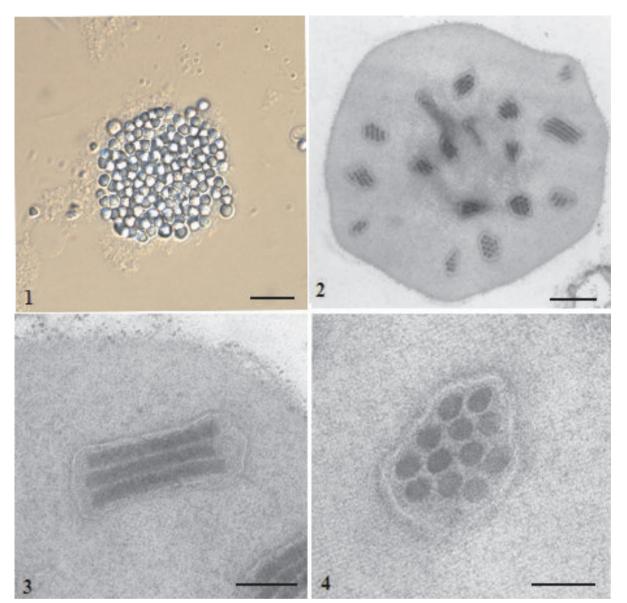
In the present study, the isolation and characterization of a novel isolate of *Leucoma salicis* nucleopolyhedrovirus (LesaNPV-TR) from Turkey are presented for the first time.

Material and Methods

Collecting infected larvae of L. salicis

Virus-infected larvae of *L. salicis* were observed on wild poplar foliage in Gümüşhane located in South Central Province of the Black sea Region of Turkey. The infected larvae were transferred to col-

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Figs 1-4. Fig. 1 – Polyhedra of *Neodiprion sertifer* nucleopolyhedrovirus, light microscope, bar = $10~\mu m$; Fig. 2 – Section of a polyhedra with virions containing multiple rod-shaped nucleocapsids, bar = 300~nm; Fig. 3 – Longitudinal section of virion, bar = 125~nm; Fig. 4 – Cross section of virion, TEM, bar = 200~nm.

lection tubes using sterile forceps. A fresh pair of forceps for each new sample was used to prevent cross-contamination of material. Collected samples were brought to the laboratory as soon as possible and stored at -20°C (HUNTER-FUJITA *et al.* 1998).

Determination of viral infection

The infected larvae were dissected, and polyhedrae were observed under a light microscope. In addition, the presence of viral infection was confirmed with Giemsa staining (HUNTER-FUJITA *et al.* 1998). Polyhedrae from the LesaNPV-TR isolate were examined by transmission (TEM) electron microscopy. Different portions of infected larvae were fixed in 2.5% glutaraldehyde in 0.1 M cacodylate buffer (pH 7.4) for 1-2 h, rinsed in

cacodylate buffer, postfixed in reduced OsO₄ according to KARNOVSKY (1971) (a fresh 1:1 mixture of 2% OsO₄ and 3% K₄ [Fe(CN₆)] for 1.5 h, rinsed in cacodylate buffer and dehydrated in an ethyl alcohol series prior to embedding in Spurr's resin (SPURR 1969). Thin sections were mounted on Pioloform-coated copper grids which were stained with saturated uranyl acetate (WATSON 1958) and Reynold's lead citrate (REYNOLD 1963). They were examined in a Philips 208 electron microscope.

Results and Discussion

The virus was detected in populations of *Leucoma salicis* in Gümüşhane. The infected larvae

were less active and dark-brown color. Their cuticula was easily broken. The majority of the dead larvae hang on poplar branches and leaves, characteristically attached with abdominal prolegs. Under the light microscope, a high number of polyhedral inclusion bodies (PIBs) formed by the virus was observed (Fig. 1). The dimension of the polyhedral inclusion bodies is $2.08\pm0.31~\mu m$ (n=50). PIBs vary in size from 1.51 to 2.64 mm. and were usually polygonal in shape (Fig. 2).

In electron microscopy studies, virions contained 2 to 15 nucleocapsids per virion (Figs 3 & 4). The size of the viral particles was 270.71±12.72 (250-290) x 33.11 ± 3.48 (32-40) nm. In a recent detailed study on the Polish isolate of LesaNPV, ZIEM-NICKA (1981) reported virions 306± 11 nm in length and 42.9±2.1 nm wide. Similarly, WEISER et al. (1954) recorded virions 270-400 x 100-150 nm in size. Clear differences are seen among Turkish, Polish and Czech isolates. As ZIEMNICKA (1981) suggests, it is rather improbable that these differences are caused by the use of various techniques and perhaps two different virus strains occur in Turkey and Poland. This idea is supported by MURILLO et al. (2001). These authors suggest that some strains of nucleopolyhedrovirus isolated from different geographic localities may possess better insecticidal activities.

LesaNPV gives promising results in the above mentioned higher insecticidal activities (HUBER 1998). Several important studies have been carried out on Leucoma salicis nucleopolyhedrovirus isolates from Western Europe (LAMERIS et al. 1985; HUBER 1998) and Eastern Europe (STROKOVS-KAYA et al. 1996; LIPA & ZIEMNICKA 1996; ZIEMNICKA 1981, 2000; JAKUBOWSKA et al. 2005). Some isolates were also recorded from different parts of Europe (WEISER et al. 1954; SKA-TULLA 1985). Unfortunately there is no record of natural isolation and characterization of this virus from Turkey as a different geographical locality from Asia, whereas Asia is a potential source of new and interesting NPV strains (MURILLO et al. 2001). It is known that some strains of nucleopolyhedrovirus isolated from different geographic localities may present better insecticidal activities, which make them more suitable for host control and show important differences in biological activity (MURILLO et al. 2001). For example, a Polish NPV isolate was about seven times more infectious than one from Yugoslavia (ZIEMNICKA 1981; LAMERIS *et al.* 1985; LIPA 1998).

In the present study, the isolation and characterization of a novel isolate of *Leucoma salicis* nucleopolyhedrovirus (LesaNPV-TR) from Turkey is presented for the first time. The results of this study will encourage scientists to compare the infectivity and systematics of LesaNPV isolates

from different parts of Europe and Asia at the molecular level, especially Polish and Turkish isolates, in order to find the most effective strain and understand their evolution.

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References

ÇANAKÇIOĞLU H. 1983. Forest Entomology, İstanbul Üniversitesi Yayınları, İstanbul. (In Turkish).

GREATHEAD D. J. 1976. A review of biological control in Western and Southern Europe. Commonwealth Agricultural Bureaux Farnham Royal, Slough, England.

JAKUBOWSKA A., OERS M. M., CORY J. S., ZIEMNICKA J., VLAK J. M. 2005. European *Leucoma salicis* NPV is closely related to North American *Orgyia pseudotsugata* MNPV. J. Invertebr. Pathol. **88**: 100-107.

HUBER J. 1998. Western Europe. (In: Insect Viruses and Pest Management. R. F. Hunter-Fujita, P. F. Entwistle, H. F. Evans, N. E. Crook eds, John Wiley & Sons, London): 201-215.

HUNTER-FUJITA R. F., ENTWISTLE P. F., EVANS H. F., CROOK N. E. 1998. General laboratory practice. (In: Insect Viruses and Pest Management. R. F. Hunter-Fujita, P. F. Entwistle, H. F. Evans, N. E. Crook eds, John Wiley & Sons, London): 359-473.

KARNOVSKY M. J. 1971. Use of ferrocyanide-reduced osmium tetroxide in electron microscopy, Proc. 14th Ann. Meet. Am. Soc. Cell Biol., p. 146.

LAMERIS A. M. C., ZIEMNICKA J., PETERS D., GRIJPMA P., VLAK J. M. 1985. Potential of baculoviruses for control of satin moth, *Leucoma salicis* L. Med. Fac. Landbouww. Rijksuniv. Gent **50**: 431-439.

LANGOR D. W. 1995. Satin moth. Nat. Resour. Can., Can. For. Serv., North. For. Cent., Edmonton, Alberta. For. Leafl. 35.

LIPA J. J. ZIEMNICKA J. 1996. Spiroplasma and microsporidian infections in populations of *Leucoma salicis* L. (Lepidoptera: Lymantriidae) in Poland. (In: Proc. Intern. Conf. "Integrated Management of Forest Lymantriidae", Warsaw-Sękocin, Poland): 27-29.

LIPA J. J. 1998. Eastern Europe and the Former Soviet Union. (In: Insect Viruses and Pest Management. R. F. Hunter-Fujita, P. F. Entwistle, H. F. Evans, N. E. Crook eds, John Wiley & Sons, London): 216-231.

MURILLO R., MUNOZ D., LIPA J. J., CABELLERO P. 2001. Biochemical characterization of three nucleopolyhedrovirus isolates of *Spodoptera exigua* and *Mamestra brassicae*. J. Appl. Ent. 125: 267-270.

REYNOLD E. S. 1963. The use of lead citrate at high pH as an electron-opaque stain in electron microscopy. J. Cell Biol. 17: 208-212.

SKATULLA U. 1985. Untersuchungen zur Wirkung eines Kernpolyedervirus aus *Leucoma salicis* (Lep., Lymantrii-

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dae) auf einige Lymantriiden- Arten. Anz. für Schändlings. Pflanzenschutz Umweltschutz. **58**: 41-47.

- SPURR A. R. 1969. A low-viscosity epoxy resin embedding medium for electron microscopy. Clin. Microbiol. Res. 3: 197-218.
- STROKOVSKAYA L. ZIEMNICKA J., MICHALIK J. 1996. Genetic variability of four natural isolates of the *Stilpnotia salicis* multiple-envoleped nuclear polyhedrosis virus. Acta Biochimica Polonica **43**: 633-638.
- WATSON M. L. 1958. Staining of tissue section for electron microscopy with heavy metals. J. Biophys. Biochem. Cytol. 4: 475.
- WEISER J., LUDVIK J., VEBER J. 1954. Polyedrie bekyne vbrove (*Stilpnotia salicis* L., Lepidoptera). Zool. Entomol. Listy 4: 238-242. (In Czech).
- ZIEMNICKA J. 1981. Studies on nuclear and cytoplasmic polyhedrosis viruses of the satin moth (*Stilpnotia salicis* L.) (Lepidoptera, Lymantriidae). Prace Nauk. Inst. Ochr. Roslin 23: 75-142.
- ZIEMNICKA J. 2000. The role of baculovirus SsMNPV in regulation numbers of the satin moth population *Stilpnotia salicis* L. (Lepidoptera: Lymantriidae). Rozprawy Naukowe Instytutu Ochrony Roślin **6**: 85 pp. (In Polish).