



Diagnoses and remarks on the genera of Tortricidae (Lepidoptera). Part 6. Grapholitini

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Abstract. Comparative diagnoses, redescrptions, and remarks are presented on the genera of the tribe Grapholitini. Original references, type species, synonyms, numbers of known species, and zoogeographic regions are provided.

Key words: Lepidoptera, Tortricidae, Grapholitini, genera, comparative diagnoses, comments.

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I. INTRODUCTION

The number of genera of Tortricidae has increased dramatically over the last 50 years; by 2007 there were over 1630 described genera, including synonyms. Many of the older descriptions are scattered throughout the literature, and because there are few larger synthetic treatments of the tortricids for most major biogeographic regions, this large number of taxa complicates considerably the work of taxonomists on the faunas of poorly known regions of the planet. In addition, characters that define many of the genera are not clearly articulated. The distribution of many genera is still insufficiently known, and this shortcoming frequently results in unexpected findings, e.g., the discovery of Afrotropical genera in the Neotropics. These types of discoveries may cause confusion for specialists that focus on the fauna of a single geographic region.

The literature abounds with re-descriptions and diagnoses of tortricid genera, but many are rather short, frequently lacking comparisons with similar or related taxa. Detailed comparative diagnoses are not only useful in systematic work but are re-

quired by the International Code of Zoological Nomenclature (1999) for descriptions of new taxa.

In this series of papers on the tortricid genera, diagnoses are based on features provided in the original description, augmented by comments from subsequent papers. My own diagnoses are proposed when no earlier ones are available. Other characteristics of the genera are included when necessary or relevant.

Morphological features that define many genera require revision and/or augmentation. Also, definitions of some genera require brief comments. Some original diagnoses are quoted verbatim, especially when no subsequent evaluation has been done. On the other hand, original and/or older diagnoses are occasionally omitted because of their limited importance.

The goal of this series of papers is to present a compilation of the existing data on tortricid genera and to identify what is known and where information is incomplete or lacking.

The account for each genus consists of the original reference, type-species (t. sp.) with the countries of origin (in case of large countries also with

their provinces, or large islands), the number of species included originally (e.g., monotypic), and the number of species known at present, the latter often based on the catalogue by BROWN (2005). The acronyms of the zoogeographic regions are added. The synonymies are treated in a similar way. The references refer to re-descriptions and diagnoses. The genera are arranged alphabetically which simplifies the index to include only synonyms.

The parts of this series are published in non-systematic order, depending on the sequence of completion of each group. The parts already published are: RAZOWSKI (2009) treating Phricanthini, Tortricini, and Schoenotenini; RAZOWSKI (2011b) treating Cochylini; RAZOWSKI (2015b) treating Archipini; and RAZOWSKI (2016) treating Cnephasiini, Ceracini, Atteriini, Sparganothini and Euliini.

Abbreviations for the zoogeographic regions are as follows:

AFR = Afrotropical, AU = Australian, HOL = Holarctic, NEA = Nearctic, NEO = Neotropical, OR = Oriental, PAL = Palearctic.

Other abbreviations are as follows: S = sternite, T = tergite, t. sp. = type species, t. l. = type locality.

II. DIAGNOSES

Acailandica RAZOWSKI & BECKER, 2016

Acailandica RAZOWSKI & BECKER, 2016, Zootaxa, **4066**(3): 2491 t. sp.: *Acailandica acailandiae* RAZOWSKI & BECKER, 2016, Brazil. Seven species included. **NEO**.

D i a g n o s i s. Originally (RAZOWSKI & BECKER, 2016), the genus was compared to *Satronia*; the latter has a weakly developed uncus with two long setae or an elongate top of the tegumen. *Satronia* differs from *Ricula* in having completely reduced socii. *Acailandica* has a telochromatic colouration of forewings resembling those of *Hilarographa* ZELLER, 1877, *Hilarographini* while the species of *Satronia* and *Ricula* have cryptic colouration.

Acanthoclita DIAKONOFF, 1982

Acanthoclita DIAKONOFF, 1982, Zool. Verh. Leiden, **193**: 27; t. sp.: *Eucosma balanopycha* MEYRICK, 1910, India. Twelve species included. **PAL, AFR, OR, AU**.

Mesotes DIAKONOFF, 1988, Annls Soc. Ent. Fr. (N.S.), **24**(2): 172; t. sp.: *Mesotes pectinata* DIAKONOFF, 1988, Madagascar. Two species included. **AFR**.

Mesotis DIAKONOFF, 1988, Annls Soc. Ent. Fr. (N.S.), **24**(2): 172 – misspelling of *Mesotes*.

Redescriptions. KOMAI & HORAK (2006), NEDOSHIVINA (2013).

D i a g n o s i s. There is no original comparative diagnosis of *Acanthoclita*; DIAKONOFF (1982) mentioned only that it: “characterizes by eucosmine venation, the spining of the clavate top of the valva and the sclerotic sterigma”.

KOMAI & HORAK (2006) compared *Acanthoclita* to *Matsumuraes* and treated it as the sister group. They share “a path of dense, modified scales usually on both sides of the hindwing between CuP and 1A+2A...”.

DIAKONOFF (1988a) stated that *Mesotes* is “allied structurally to *Leguminivora*. *Mesotes* differs from *Leguminivora* by the male genitalia, in the former rather resembling those of *Grapholita*, while in the latter they approach to the male genitalia of *Fulcrifera* FALKOVITCH, another true grapholitine”.

Age DIAKONOFF, 1982

Age DIAKONOFF, 1982, Zool. Verh. Leiden, **193**: 56; t. sp.: *Age onychistica* DIAKONOFF, 1982, Sri Lanka. Three species included. **OR, AFR**.

D i a g n o s i s. According to DIAKONOFF (1982) *Age* is “apparently allied to *Acanthoclita* gen. nov., with the venation almost congruent, but with a quite different wing shape and the course of the media in the cell of the fore wing, and with characteristic male genitalia”.

Agriophanes MEYRICK, 1930

Agriophanes MEYRICK, 1930, Exotic Microlepid., **3**: 600; t. sp.: *Agriophanes pycnostrota* MEYRICK, 1930, India: Madras. One species included. **OR**.

D i a g n o s i s. MEYRICK (1930) stated that this genus show “characters of *Argyroploce* [Olethreutini], but hindwings 3 and 4 coincident. May rank next to *Helictophanes*”.

KOMAI (1980) regarded *Agriophanes* as related to *Pseudopammene* and *Dierlia*. KOMAI (1999) concluded these genera are closely related, all “sharing long-stalking or the coincidence of M3 and CuA1 in the hindwing and the ductus bursae with an ovate sclerite with a concavity”.

R e m a r k s. Also mentioned under *Pseudopammene*.

Andinarampha HEPPNER, 2013

Andinarampha HEPPNER, 2013, Lepid. Novae, **6**: 47; t. sp.: *Andinarampha nanoflava* HEPPNER, 2013, India: Loja. Four species included. **NEO**.

D i a g n o s i s. According to HEPPNER (2013b), *Andinarampha* is related to *Satronia* in general appearance and in a few morphological details such as the absence of the hindwing cubital pecten and lack of socii in the male genitalia. The maculation differs from that of other Grapholitini genera in the

absence of costal strigulae and the forewing fascia uninterrupted from the costa to the dorsum.

Andrioplecta OBRAZTSOV, 1968

Andrioplecta OBRAZTSOV, 1968, J. New York Ent. Soc., **76**: 176; t. sp.: *Laspeyresia pulverula* MEYRICK, 1912, India: Assam. Ten species included. **PAL, OR, AU**.

Redescription. KOMAI (1992, 1999).

Diagnosis. OBRAZTSOV (1968) described *Andrioplecta* as a probable “development of *Laspeyresia* HÜBNER. As to the venation, the new genus approaches *Strophedra* HERRICH-SCHÄFFER, but differs from it by having veins M_3 and Cu_1 in the forewing approximated at termen, and strong sclerotization of the dorsum in the hindwing of the male...”.

KOMAI (1992) included *Andrioplecta* to the *Grapholita-Pammene* complex which consists of ten genera and suggested that it “may be most closely allied to *Strophedra*...”. KOMAI (1999) later concluded that *Andrioplecta* is closest to *Strophedra* based on the presence of “flap structures” of the tegumen.

Remarks. Also mentioned under *Apocytia*, *Cydia*, *Ixonympha*, *Leguminivora* and *Parapammene*.

Apocytia KOMAI & HORAK, 2006

Apocytia KOMAI & HORAK, 2006, Monogr. Aust. Lepid., **10**: 439; t. sp.: *Eucosma pervicax* MEYRICK, 1911, Australia: North Territory. One species included. **AU**.

Diagnosis. KOMAI & HORAK (2006) compared *Apocytia* to *Cydia*; both have a “concave ventral sacculus margin near its base... and a modified hindwing anal margin, albeit only thickened in *Apocytia* and not folded as in *Cydia*. The thickened male antenna, the shape of the valva, the cup or pocket-shaped sterigma and the twisted ductus bursae are autapomorphies for *Apocytia*”.

Archiphlebia KOMAI & HORAK, 2006

Archiphlebia KOMAI & HORAK, 2006, Monogr. Aust. Lepid., **10**: 433; t. sp.: *Argyroploce endophaga* MEYRICK, 1911, Australia: Western Australia. Two species included. **AU**.

Diagnosis. According to the original diagnosis *Archiphlebia* is related to *Thaumatotibia*, *Cryptophlebia* and *Gymnandrosoma* but differs from them by two apomorphies: “the posterior half of ductus bursae narrow and curved near middle, and S_7 with unscaled medio-anterior region”.

Articolla MEYRICK, 1907

Articolla MEYRICK, 1907, J. Bombay Nat. Hist. Soc., **17**: 976; t. sp.: *Articolla cyclidias* MEYRICK, 1907, Sri Lanka. One species included. **OR**.

Diagnosis. MEYRICK (1907) stated: “Allied to *Platyephus* [= *Dudua*, *Olethreutini*] from which it differs by the stalking of 8 and 9 of forewing”.

Balbis WALSINGHAM, 1897

Balbis WALSINGHAM, 1897, Proc. Zool. Soc. London, **1897**: 128; t. sp.: *Carpocapsa assumptana* WALKER, 1863, Brazil: Amazonas. One species included. **NEO**.

Redescription. RAZOWSKI (2011a).

Diagnosis. WALSINGHAM (1897) stated: “Agreeing with *Dichrorampha* and *Lipoptycha* in having veins 6 and 7 in hind wings parallel, but differing in the palpi and in the unsinate termen, and probably intermediate between these genera and *Laspeyresia*, HB. (= *Grapholitha* HS.)”.

HEINRICH (1926) compared *Balbis* to *Talponia*.

RAZOWSKI (2011a) wrote that *Balbis* is probably distinct from *Dichrorampha* and is either closely related to it, or is a senior synonym of *Ricula*. The main difference between the two is the presence of a very long, apomorphic basal process of the valva in *Balbis*. He also compared *Balbis* to *Ricula*.

Remarks. Also mentioned under *Archiphlebia* and *Talponia*.

Celsumaria BROWN & TIMM, 2017

Celsumaria BROWN & TIMM, 2017, Anns Ditsong Mus. Nat. Hist., **7**: 98; t. sp.: *Celsumaria knysna* BROWN & TIMM, 2017, South Africa. Five species known. **AFR**.

Diagnosis. *Celsumaria* is related to *Thaumatotibia*, but *Celsumaria* has an orbicular organ near the middle of the disc of the valva connected by a rounded opening to the outer surface of the valva. A similar and probably homologous structure is found also in the *niphadonta*-group of *Dracontogena* and in *Thylacandra*. In facies, *Celsumaria* is similar to *Cryptasasma* WALSINGHAM, 1900, *Microcorsini* and *Dracontogena*. According to the original description, *Celsumaria* differs from these genera in having “small rounded scales of raised, ribbon-like scales”. Based on facies and male genitalia, *Celsumaria* is most likely a junior synonym of *Thylacandra*.

Mentioned also with *Cryptasasma*, *Dracontogena* and *Thaumatotibia*.

Centroxena DIAKONOFF, 1971

Centroxena DIAKONOFF, 1971, Veröff. Zool. Staatsmus. München, **15**: 182; t. sp.: *Centroxena ulophora* DIAKONOFF, 1971, Thailand. One species included. **OR, AU**.

Diagnosis. DIAKONOFF (1971) – compared *Centroxena* to *Goditha* which have similar male genitalia “but without doubt, only superficially, because of many other structural differences, as e.g. the stalked veins 3 and 4 of the modified

hindwing. Also somewhat resembling *Collogenes* MEYRICK, 1931 [Microcorsini], but easily separable by the separate veins 7 and 8 of the fore wing”.

Coccothera MEYRICK, 1914

Coccothera MEYRICK, 1914, Ann. Transvaal Mus., **4**: 189; t. sp.: *Grapholitha spissana* ZELLER, 1952, South Africa: Natal. Nine species included. **PAL, AFR.**

Cirriphora OBRAZTSOV, 1951, Tijdschr. Ent., **93**(1950): 99; t. sp.: *Grapholitha pharaonana* KOLLAR, 1858, Egypt.

Endotera AGASSIZ, 2011, J. Nat. Hist., **45**(29-30): 1887; t. sp.: *Endotera nodi* AGASSIZ, 2011, Kenya. **Syn. n.**

Redescriptions. DANILEVSKY & KUZNETZOV (1968, *Cirriphora*), DIAKONOFF (1968, *Coccothera*), RAZOWSKI (1989, 2004, 2015a, *Coccothera*).

Diagnosis. MEYRICK (1914) stated: “The examination... shows that this species [*Grapholitha spissana*] forms a new genus, allied to *Laspeyresia*”.

OBRAZTSOV (1951) characterized *Cirriphora* as follows: “A monotypic genus, probably mediterranean in origin. Belongs to the group of *Gymnandrosoma* DYAR and *Ecdytophaga* Z. but differs from them in the genitalia and in the presence of androconial pencil in hindwing of male”.

AGASSIZ (2011) compared *Endotera* to *Eucosma* (Eucosmini) as having similar venation but the former with “the ”flap“ of membrane of the inside of the valva is a characteristic of the genus, together with the sclerotization on tergite 8 of the female”.

Remarks. The genitalia of *Endotera nodi* are very similar to those of *Cirriphora pharaonana* and *Grapholitha spissana* which according to DIAKONOFF (1968) differs from his *Coccothera ferrifracta* only in markings. A discussion of the synonymy of *Cirriphora* can be found in RAZOWSKI (2015a).

Commoneria KOMAI & HORAK, 2006

Commoneria KOMAI & HORAK, 2006, Monogr. Aust. Lepid., **10**: 459; t. sp.: *Laspeyresia cyanosticha* TURNER, 1946, Australia: Queensland. One species included. **AU.**

Diagnosis. KOMAI & HORAK (2006) compared *Commoneria* to *Microsarotis*. The two have “the dorsal scent organ associated with T8 and also R_s and M1 distant at base, but the genitalia do not suggest a close relationship between the two genera. Sternum 8 is as long as T8 and with a concave hind margin that is unusual among Grapholitini except for *Loranthacydia*”.

Remarks. Also mentioned under *Microsarotis*.

Coniostola DIAKONOFF, 1961

Coniostola DIAKONOFF, 1961, Anns Soc. Ent. Fr., **130**: 71; t. sp.: *Eucosma stereoma* MEYRICK, 1912, India: Bengal. Eight species included. **OR, AFR.**

Diagnosis. There is no original comparative diagnosis; DIAKONOFF (1961) mentioned only that “*Coniostola* undoubtedly belongs to the *Laspeyresiini*”.

Remarks. The male genitalia of *Coniostola* have the valvae similar to those of many *Grapholitha* species but have a strongly broadened proximal part of the aedeagus. The female genitalia resemble those of *Lathronympha* but have a ring-shaped sclerite at the base of the ductus bursae and a proximal, broad basal part of the ducus seminalis.

Corticivora CLARKE, 1951

Corticivora CLARKE, 1951, J. Wash. Acad. Sci., **41**: 46; t. sp.: *Corticivora clarki* CLARKE, 1951, USA: Connecticut. Three species included. **PAL, NEA.**

Redescriptions. BROWN (1984) MILLER (1987), GILLIGAN et al. (2008).

Diagnosis. CLARKE (1951) compared *Corticivora* to *Gypsonoma* MEYRICK, 1895 finding some differences in their wing venation but stated that it “appears to be most nearly related to *Laspeyresia* but differs from it by the stalking of veins 6 and 7 of the hindwing, the presence of socii, and the form of the signa”. BROWN (1984) wrote that “a comprehensive study of the world fauna of ”Grapholitini“ is needed to resolve ancestral relationships and to associate the currently included genera with their sister groups. BROWN (2005) included four species in *Corticivora* (also the Palaearctic *Tortrix* (*Coccyx*) *piniana* HERRICH-SCHÄFFER). RAZOWSKI (2003) followed OBRAZTSOV’s (1964) interpretation and retained *Corticivora* in Eucosmini.

Cryptophlebia WALSINGHAM, 1899

Cryptophlebia WALSINGHAM, 1899, Indian Mus. Notes, (1899)**4**(3): 105; t. sp.: *Cryptophlebia carpophaga* WALSINGHAM, 1899 = *Arothrophora ombrodelta* LOWER, 1898, Australia: New South Wales. Fifty species included. **PAL, OR, AFR, AU.**

Pogonozada HAMPSON, 1905, Ann. Mag. nat. Hist., (7)**16**: 586; t. sp.: *Pogonozada distorta* HAMPSON, 1905, China: Ichang.

Phanerophlebia DIAKONOFF, 1957, Tijdschr. Entomol., **100**: 142; t. sp.: *Cryptophlebia perfracta* DIAKONOFF, 1957, Indonesia: Java.

Redescriptions. BRADLEY (1953), RAZOWSKI (1989, 2004), KOMAI (1999), KOMAI & HORAK (2006), NEDOSHIVINA (2013).

Diagnosis. According to KOMAI (1999) *Cryptophlebia* “has some relationships with *Thaumatotibia* ZACHER and the North American *Ecdy-*

tolopha ZELLER and *Pseudogalleria* RAGONOT...". KOMAI (1999) wrote that "monotypy of *Cryptophlebia* is supported by (1) T8 subtriangular or Y-shaped with a pair of tufts of long filiform scales arising from shallow membranous pockets on the posterior edges, (2) 8th sternite of the coremata with a pair of short projections laterally, (3) thickly swollen, clavate valva with sparse strong spines on the inner surface of the cucullus, and (4) corpus bursae aciculate in anterior 1/2-4/5". The monotypic, North American genus *Pseudogalleria* has no important characters that differ from *Cryptophlebia*, and it most likely is a senior synonym of the latter.

Remarks. RAZOWSKI (1989, 2004) mentioned putative autapomorphies for *Cryptophlebia*.

Cryptophlebia is also mentioned under *Coccolthera*, *Cryptoschesis*, *Dracontogena*, *Ecdytolopha*, *Gymnandrosoma*, *Matsumuraeses*, *Pseudogalleria*, *Thaumatotibia*, and *Thylacandra*.

Cryptoschesis DIAKONOFF, 1988

Cryptoschesis DIAKONOFF, 1988, *Annls Soc. Ent. Fr. (N.S.)*, **24**(3): 318; t. sp.: *Cryptoschesis imitans* DIAKONOFF, 1988, Madagascar. One species included. **AFR**.

Diagnosis. Originally (DIAKONOFF, 1988b) compared *Cryptoschesis* to *Cryptophlebia* from which it differs by the "grapholite vinculum and by the attachment of the valvae in the present genus; these parts are decidedly more eucosmine in the new genus".

Cyanocydia RAZOWSKI & BECKER, 2012

Cyanocydia RAZOWSKI & BECKER, 2012, *Polish J. Entomol.*, **81**(3): 204; t. sp.: *Cydia eucyanea* WALSINGHAM, 1914, Mexico: Veracruz. Three species included. **NEO**.

Diagnosis. Externally, RAZOWSKI & BECKER (2012) compared *Cyanocydia* to *Cydia*. The male genitalia are distinguished by the very large, densely scaled socii and the female genitalia somewhat resembling those of the genera of the *Dichrorampha*-group having the sterigma included in the subgenital sternite. The putative autapomorphies for *Cyanocydia* are the strongly sclerotized rod of the sterigma fused with the posterior edge of the sternite and a very long postostial sterigma.

Remarks. Two species described by HEPPNER (2013a) in *Cydia* belong in *Cyanocydia* and are transferred herein: *Cyanocydia salvadorana* (HEPPNER, 2013) and *Cyanocydia costaricensis* (HEPPNER, 2013).

Cydia HÜBNER, [1825]

Cydia HÜBNER, [1825], *Verz. bekannter Schmett.*: 375; t. sp.: *Phalaena pomonella* LINNAEUS, 1758, Europe. Over 220 species included. **PAL**, **NEA**, **NEO**, **AU**.

Laspeyresia HÜBNER, [1825], *Verz. bekannter Schmett.*: 375; t. sp.: *Tortrix corollana* HÜBNER, [1823], Europe. Junior homonym of *Laspeyresia* R. L., 1817.

Erminea KIRBY & SPENCE, 1826, *Introd. Entomol.*, **3**: 123; t. sp.: *Phalaena pomonella* LINNAEUS, 1758, Europe. Junior homonym of *Erminea* HAWORTH, [1811], Lepidoptera, Yponomeutidae.

Carpocapsa TREITSCHKE, 1829, [in] OCHSENHEIMER, *Schmett. Eur.*, **7**: 230; t. sp.: *Phalaena pomonella* LINNAEUS, 1758, Europe.

Coccyx TREITSCHKE, 1829, [in] OCHSENHEIMER, *Schmett. Eur.*, **7**: 230; t. sp.: *Tortrix strobilana* HÜBNER, [1799], Europe.

Semasia STEPHENS, 1829, *Nom. Br. Insects*: 47; t. sp.: *Phalaena pomonella* LINNAEUS, 1758, Europe.

Strobila SODOFFSKY, 1837, *Bull. Soc. Imp. Nat. Moscou*, **1837**(6): 92. Junior homonym of *Strobila* SARS, 1829, Coleoptera and unnecessary replacement for *Coccyx*.

Carpocampa HARRIS, 1841, *Rep. Insects Mass. injurious to Vegn.*: 351. Emendation of *Carpocapsa* TREITSCHKE, 1829.

Cerata STEPHENS, 1852, *List. Specimens Br. Anim. Colln Br. Mus.*, **10**: 77; t. sp.: *Penthina servillana* DUPONCHEL, 1836, France.

Melissopus RILEY, 1882, *Trans. Acad. Sci. St. Louis*, **4**: 322; t. sp.: *Carpocapsa latiferreana* WALSINGHAM, 1879, USA: California, Oregon.

Melissopus RILEY, 1882, *Trans. Acad. Sci. St. Louis*, **4**: 322 – misspelling of *Melissopus*.

Melissopus FERNALD, 1882, *Trans. Am. Ent. Soc.*, **10**: 54 – misspelling of *Melissopus*.

Melliopus PACKARD, 1890, *Fifth Rep. U.S. Ent. Comm.*: 219 – misspelling of *Melissopus*.

Melissopus FERNALD, 1908, *Genera Tortricidae Types*: 60 – misspelling of *Melissopus*.

Adenoneura WALSINGHAM, 1907 [in] SHARP, *Fauna Hawaii*, **1**(5): 677; t. sp.: *Adenoneura falsifalcellum* WALSINGHAM, 1907, Hawaii.

Crobylophora KENNEL, 1908, *Zool. Stuttg.*, **21**(54): 50; t. sp.: *Tortrix inquinatana* HÜBNER, [1796-1799], Europe. Junior homonym of *Crobylophora* MEYRICK, 1880, preoccupied.

Hedulia HEINRICH, 1926, *U.S. Natn. Mus. Bull.*, **132**: 6; t. sp.: *Hedulia injectiva* HEINRICH, 1926, USA: Nevada.

Kenneliola PACLT, 1951, *Revue fr. Lepid.*, **13**: 127; t. sp.: *Tortrix inquinatana* HÜBNER, [1796-1799], Europe. Objective replacement name for *Crobylophora* KENNEL.

Pseudotomoides OBRAZTSOV, 1959, *Tijdschr. Ent.*, **102**: 200; t. sp.: *Phalaena strobilella* LINNAEUS, 1758, Europe.

Collicularia OBRAZTSOV, 1960, *Tijdschr. Ent.*, **103**: 60; t. sp.: *Catoptria microgrammana* GUENÉE, 1845, France.

Phanetoprepa OBRAZTSOV, 1968, *J. New York Ent. Soc.*, **76**: 236; t. sp.: *Phanetoprepa agenjoi* OBRAZTSOV, 1968, Spain.

Danilevskia KUZNETZOV, 1970, *Entomol. Obozr.*, **49**: 446; t. sp.: *Danilevskia silvana* KUZNETZOV, 1970, Russia: Primorsky Krai.

Dicraniana DIAKONOFF, 1984, *Entomol. Gall.*, **1**: 162; t. sp.: *Semasia seriana* KENNEL, 1901, Spain. Proposed as a subgenus of *Cydia*.

Redescriptions. KENNEL (1921, as *Laspeyresia*), HEINRICH (1926 as *Laspeyresia*), OBRAZTSOV (1959 as *Laspeyresia*, also *Collicularia*, *Pseudotomoides*), DANILEVSKY & KUZNETZOV (1968 as *Laspeyresia*), MILLER (1987),

RAZOWSKI (1989, 2003, 2004), KOMAI (1999, phylogeny discussed), KOMAI & HORAK (2006), GILLIGAN et al. (2008), RAZOWSKI & BROWN (2012).

Diagnosis. HEINRICH (1926) compared *Cydia* to *Grapholita* and its synonyms *Hedulia* (the hairy vestiture is unique in the family and is reminiscent of *Synnoma* [Sparganothini]) and *Melissopus* to *Cydia*. *Carpocapsa* was also preserved as a distinct genus.

DANILEVSKY & KUZNETZOV (1968) compared *Cydia* to *Fulcrifera*, *Leguminivora*, *Lathronympha* and *Cirriphora* and placed it between *Leguminivora* and *Lathronympha*.

KOMAI & HORAK (2006) compared *Cydia* to *Leguminivora*, *Notocydia*, *Fulcrifera*, and *Apocydia*.

Remarks. According to KOMAI (1999) and KOMAI & HORAK (2006), the *Cydia*-group of genera shares three characters: ventral margin of sacculus concave near base, presence of anal fold of male hindwing, and vein 3A close to anal edge of male hindwing.

DANILEVSKY & KUZNETZOV (1968) divided *Laspeyresia* (= *Cydia*) into three subgenera: *Endopisa*, *Laspeyresia*, and *Kenneliola*. RAZOWSKI (2003) and KOMAI & HORAK (2006) have not distinguished the subgenera.

Cydia is also mentioned under *Apocydia*, *Balbis*, *Coccothera*, *Cyanocydia*, *Dichrorampha*, *Eriosocia*, *Grapholita*, *Karacaoglania*, *Larisa*, *Lathronympha*, *Macrocydia*, *Metacydia*, *Multiquestia*, *Pammenemima*, and *Phloerampha*.

Dichrorampha GUENÉE, 1845

Dichrorampha GUENÉE, 1845, Annl. Soc. Ent. Fr., (2)3: 185; t. sp.: *Grapholita plumbagana* TREITSCHKE, 1830, Austria. Ca. 150 species included. **PAL**, **NEA**, **NEO**.

Amaurosetia STEPHENS, 1835, Illustr. Br. Ent., 4, Haustelata: 353; t. sp.: *Phalaena albinella* LINNAEUS, 1758 = *Elachista megerlella* HÜBNER, 1810, Europe.

Dichroramphodes OBRAZTSOV, 1953, Mitt. Münch. ent. Ges., 43: 77; t. sp.: *Dichrorampha gueneana* OBRAZTSOV, 1953 = *Dichrorampha vancouverana* MCDUNNOUGH, 1935, Canada: Vancouver Island.

Dichrorampha FREY, 1880, Lepid. Schweiz: 330 – misspelling for *Dichrorampha*.

Dicrorampha DOUBLEDAY, 1850, Synon. List Br. Lepid.: 26 – misspelling for *Dichrorampha*.

Lipoptycha LEDERER, 1859, Wien. Ent. Monatschr., 3: 370; *Coccyx bugnionana* DUPONCHEL, 1842, France.

Lepidoptycha DYAR, 1901, Proc. Ent. Soc. Wash., 4: 469 – incorrect subs. spelling of *Lipoptycha*.

Lipoptychodes OBRAZTSOV, 1953, OBRAZTSOV, 1953, Mitt. Münch. ent. Ges., 43: 60; t. sp.: *Coccyx bugnionana* DUPONCHEL, 1842, France – subgenus of *Dichrorampha*.

Paralipoptycha OBRAZTSOV, 1958, Tijdschr. Ent., 101: 244; t. sp.: *Phalaena plumbana* SCOPOLI, 1763, Slovenia – nom. n. for *Lipoptycha* HEINRICH, 1926, subgenus of *Dichrorampha*.

Redescriptions. HEINRICH (1926), KENNEL (1921, as *Hemimene*), OBRAZTSOV (1953, 1958), DANILEVSKY & KUZNETZOV (1968), MILLER (1987), RAZOWSKI (2003, 2011a), GILLIGAN et al. (2008).

Diagnosis. OBRAZTSOV (1958) compared the wing venation of *Dichrorampha* to that of several genera (eg. *Goditha*, *Satronia*), and the scent scales of the forewing costal fold to *Cirriphora*, and some genital characters to those of *Enarmonia*, *Pseudophiaris* and others.

KENNEL (1921) separated *Lipoptycha* from *Hemimene* by its lack of the costal fold in the male forewing.

RAZOWSKI (2011a) compared the male genitalia of *Dichrorampha* to those of *Cydia* and *Grapholita*, all sharing an expanded distal part of the valva.

Remarks. OBRAZTSOV (1953) divided *Dichrorampha* into three subgenera, *Lipoptycha*, *Dichroramphodes*, and *Dichrorampha* s. str., and in 1958 added the subgenus *Paralipoptycha* on basis of a lack of the forewing costal fold of males. DANILEVSKY & KUZNETZOV (1968) distinguished two subgenera, *Lipoptycha* and *Dichrorampha* s. str., and RAZOWSKI (1989, 2003) rejected the subdivision.

Dichrorampha is also mentioned under *Balbis*, *Eriosocia*, *Ethelgoda*, *Goditha*, *Microsarotis*, *Pammenemima*, *Phloerampha*, *Ranapoaca*, *Ricula* and *Riculomorpha*.

Dierlia DIAKONOFF, 1976

Dierlia DIAKONOFF, 1976, Zool. Verh. Leiden., 144: 30; t. sp. *Dierlia aurata* DIAKONOFF, 1976, Nepal. Two species included. **OR**.

Redescription. KOMAI (1999).

Diagnosis. According to DIAKONOFF (1976), *Dierlia* is “superficially nearest to *Pammene* HÜBNER, except for the peculiar androconial field upon the hind wing and the stalking or coincidence of veins 3 and 4 in the hind wing in the both sexes... shows a close affinity with the genus *Parapammene*... From the last genus, *Dierlia* can be separated at once by the absence of any coremata.... The genus may be allied to *Diamphidia* OBRAZTSOV, but is differing by the absence of vein 4 in the hind wing of the two sexes”.

According to KOMAI (1999), the apomorphies distinguishing the genus from *Pseudopammene* are (1) a large androconial field consisting of thin grey hair-like scales on dorsal surface of hindwing and (2) a transverse patch of dark scales on T6 of male abdomen."

***Dracontogena* DIAKONOFF, 1970**

Dracontogena DIAKONOFF, 1970, Mém. ORSTOM, **37**: 122; t. sp.: *Dracontogena niphodonta* DIAKONOFF, 1970, Madagascar. Ten species included. **AFR**.

Redescription. KARISCH (2005)

Diagnosis. According to DIAKONOFF (1970) male genitalia are of *Cryptophlebia* type and the venation of the hind wing are similar to that genus; pattern and colouration resemble those of *Hermenias* MEYRICK, 1911 [Eucosmini]. The unusual orbicular organ in the middle of the valva of members of the *niphadonta*-group of *Dracontogena* is shared with *Celsumaria* and *Thylacandra*.

KARISCH (2005) compared *Dracontogena* to *Cryptophlebia* and *Thaumatotibia*.

Remarks. Mentioned also under *Celsumaria*, *Cryptophlebia*, *Thaumatotibia*, and *Thylacandra*.

***Ecdytophpa* ZELLER, 1875**

Ecdytophpa ZELLER, 1875, Verh. zool.-bot. Ges. Wien, **25**: 266; t. sp.: *Ecdytophpa insiticihana* ZELLER, 1875, USA: Massachusetts. Twelve species included. **NEA, NEO**.

Ecdytophpa FRACKER, 1915, Illinois Biol. Monogr., **2**(1): 74 – misspelling.

Redescriptions. HEINRICH (1926), MILLER (1987), KOMAI (1999), ADAMSKI & BROWN (2001), GILLIGAN et al. (2008).

Diagnosis. Based on the colouration of the adult ZELLER (1875) compared *Ecdytophpa* to *Penthina* TREITHSCHKE [Olephreutini].

HEINRICH (1926) stated that *Ecdytophpa* is closely related to *Gymnandrosoma* “and with affinities to the *Endothenia* group of the Olethreutinae... The genitalia are typically Laspeyresiini; but otherwise the genus would go better with *Endothenia* than with *Laspeyresia*. Probably a primitive form and (with *Gymnandrosoma*) linking the Laspeyresiinae and Olethreutinae”.

According to KOMAI (1999) *Cryptophlebia* “has some relationships with *Thaumatotibia* ZACHER and the North American *Ecdytophpa* ZELLER and *Pseudogalleria* RAGONOT...”.

ADAMSKI & BROWN (2001) proposed a hypothesized phylogeny of the *Cryptophlebia-Ecdytophpa* group of genera and compared *Ecdytophpa* to *Thaumatotibia*, *Gymnandrosoma*, *Pseudogalleria*, and *Cryptophlebia*.

According to GILLIGAN et al. (2008) *Ecdytophpa* is similar to *Gymnandrosoma* but “the valva lacks the setose ridge on the ventral margin of the neck”.

Remarks. Also mentioned under *Coccothera*, *Cryptophlebia*, *Gymnandrosoma*, *Lusterola*, *Pseudogalleria*, and *Thaumatotibia*.

***Eriosocia* RAZOWSKI & BROWN, 2008**

Eriosocia RAZOWSKI & BROWN, 2008, Proc. Entomol. Soc. Wash., **110**(3): 636; t. sp.: *Laspeyresia guttifera* MEYRICK, 1913, Costa Rica. Two species included. **NEO**.

Diagnosis. Originally (RAZOWSKI & BROWN, 2008), *Eriosocia* was compared (a similar facies) to *Cydia*, *Dichrorampha* and in the genitalia to *Thylacogaster* (Enarmoniini). *Eriosocia* is distinct chiefly by some characters of the tegumen and the abdominal sex scales.

***Ethelgoda* HEINRICH, 1926**

Ethelgoda HEINRICH, 1926, Bull. U.S. Natn. Mus., **132**: 23; t. sp.: *Phthoroblastis texanana* WALSINGHAM, 1879, USA: Texas. Six species included. **NEA, NEO**.

Redescription. RAZOWSKI & BECKER (2012).

Diagnosis. HEINRICH (1926) wrote: “On wing pattern and general habitus it should go with *Talponia* (*T. plummeriana* and *E. texanana* differ superficially only in color). On male genitalia and abdominal characters it could go in *Goditha*. Its female genitalia (except for the two signa) are those of *Dichrorampha*. Its hind wing venation is that of *Ricula*.... Derived from *Goditha*”.

Remarks. Also mentioned under *Metacydia*, *Phloerampha*, and *Talponia*.

***Eucosmocystia* DIAKONOFF, 1988**

Eucosmocystia DIAKONOFF, 1988, Anns Soc. Ent. Fr. (N.S.), **24**(3): 326; t. sp.: *Eucosmocystia oedipus* DIAKONOFF, 1988, Madagascar. Three species included. **AFR**.

Diagnosis. Originally (DIAKONOFF, 1988b) compared the male genitalia of *Eucosmocystia* to those of *Grapholita*, not mentioning the differences.

Remarks. Also mentioned under *Matsumuraes*.

***Fulcrifera* DANILEVSKY & KUZNETZOV, 1968**

Fulcrifera DANILEVSKY & KUZNETZOV, 1968, Fauna SSSR, **5**(1): 454; t. sp.: *Laspeyresia luteiceps* KUZNETZOV, 1962, Russia: Siberia. Twenty-eight species included. **PAL, OR, AFR, AU**.

Redescriptions. RAZOWSKI (1989, 2004), KOMAI & HORAK (2006), RAZOWSKI & BROWN (2012).

Diagnosis. In the original description, DANILEVSKY & KUZNETZOV (1968) included *Fulcrifera* in the *Laspeyresia* (= *Cydia*) group of genera and mentioned that in the male genitalia it resembles *Leguminivora*, and that *Fulcrifera* is closely related to *Laspeyresia* (= *Cydia*) especially to the subgenus *Endopisa* [now in *Grapholita*].

KOMAI & HORAK (2006) compared *Fulcrifera* to *Leguminivora* (see diagnosis of latter).

Remarks. Also mentioned under *Acanthoclita*, *Amabrana*, *Cydia* and *Leguminivora*.

***Goditha* HEINRICH, 1926**

Goditha HEINRICH, 1926, Bull. U.S. Natn Mus., **123**: 8; t. sp.: t. sp., *Goditha bumeliana* HEINRICH, 1926, USA: Texas. Six species included. **NEA, NEO.**

Redescription. RAZOWSKI & BECKER (2013).

Diagnosis. In the original description (HEINRICH, 1926) wrote: "A development of *Dichrorampha*".

Remarks. Also mentioned under *Centroxena*, *Dichrorampha*, *Ethelgoda*, *Ranapoaca*, *Riculorampha* and *Sereda*.

***Grapholita* TREITSCHKE, 1929**

Grapholita TREITSCHKE, 1929, Schmett. Eur., **7**: 232; t. sp.: *Tortrix lunulana* [DENIS & SCHIFFERMÜLLER], 1775 = *Pyralis dorsana* FABRICIUS, 1775, Germany = *Phalaena petiverella* LINNAEUS, 1758, Sweden. 136 species included. **PAL, OR, NEA, NEO, AU.**

Grapholitha TREITSCHKE, 1830, [in] OCHSENHEIMER, Schmett. Eur., **8**: 203. Unjustified emendation of *Grapholita* TREITSCHKE, 1830.

Euspila STEPHENS, 1834, Illustr. Br. Entomol. (Haustellata), **4**: 103; t. sp.: *Tinea compositella* FABRICIUS, 1775, Great Britain.

Ephippiphora DUPONCHEL, 1834, Anns Soc. Ent. Fr., (2)**3**: 446; t. sp.: *Pyralis dorsana*: DUPONCHEL 1834 = *Phalaena jungiella* CLERCK, 1759, Europe.

Stigmonota GUENÉE, 1845, Anns Soc. Ent. Fr., (2)**3**: 182; t. sp.:

Phalaena jungiella CLERCK, 1759, Europe.

Endopisa GUENÉE, 1845, Anns Soc. Ent. Fr., (2)**3**: 182; t. sp.: *Grapholitha nebritana* TREITSCHKE, 1830,

Ebisma WALKER, 1866, List Specimens Lepid. Insects Colln Br. Mus., **35**: 1803; t. sp.: *Ebisma seclusana* WALKER, 1866, New Guinea.

Redescriptions. HEINRICH (1926), OBRAZTSOV (1959), DANILEVSKY & KUZNETZOV (1968), MILLER (1987), RAZOWSKI (1989, 2003, 2004), KOMAI (1999), KOMAI & HORAK (2006), GILLIGAN et al. (2008).

Diagnosis. According to HEINRICH (1926), *Grapholita* is derived from *Laspeyresia* (= *Cydia*).

DANILEVSKY & KUZNETZOV (1968) compared *Grapholita* to *Matsumuraeses* and *Parapammene*.

KOMAI (1999) does not provide a comparative diagnosis. He distinguished the subgenus *Aspila*.

Remarks. The monophyly of *Grapholita* is insufficiently supported by morphological characters of the adults. KOMAI (1999) suggested two putative autapomorphies, a pheromone component and the larval chaetotaxy. Also mentioned under

Acanthoclita, *Balbis*, *Coniostola*, *Cydia*, *Dichrorampha*, *Fulcrifera*, *Hyposarotis*, *Macrocydia*, *Matsumuraeses*, *Microsarotis*, *Parapammene*, *Selania*, *Sereda*, *Spanistoneura*, *Steganoptycha*, and *Strophedra*.

Grapholita was divided into two subgenera, *Grapholita* s. str. and *Aspila* (DANILEVSKY & KUZNETZOV, 1968; RAZOWSKI, 1989, 2003; KOMAI, 1999; KOMAI & HORAK, 2006).

Subgenus *Aspila* STEPHENS, 1834

Aspila STEPHENS, 1834, Illustr. Br. Entomol. (Haustellata), **4**: 104; t. sp.: *Phalaena lediana* HAWORTH, [1811] = *Coccyx janthinana* DUPONCHEL, 1853, France. **PAL, NEA, OR, AU.**

Opadia GUENÉE, 1845, Anns Soc. Ent. Fr., (2)**3**: 182; t. sp.: *Grapholitha funebrana* TREITSCHKE, 1835, Germany and Czech Republic.

Coptoloma LEDERER, 1859, Wien. Ent. Monatschr., **3**: 124, 370; t. sp.: *Coccyx janthinana* DUPONCHEL, 1835, France.

Redescriptions. DANILEVSKY & KUZNETZOV (1968), MILLER (1987), RAZOWSKI (1989, 2003), KOMAI & HORAK (2006).

Diagnosis. KOMAI (1999) compared *Aspila* to *Grapholita* s. str. mentioning their differences in genitalia and coremata.

***Gymnandrosoma* DYAR, 1904**

Gymnandrosoma DYAR, 1904, Proc. Entomol. Soc. Wash., **6**: 60; t. sp.: *Gymnandrosoma punctidiscanum* DYAR, 1904, USA: District Columbia. Eight species included. **NEA, NEO, AU.**

Gynandrosoma SHARP, 1905, Zool. Record, **41**, Insecta: 291 – misspelling.

Redescriptions. HEINRICH (1926), ADAMSKI & BROWN (2001, revision), KOMAI & HORAK (2006), GILLIGAN et al. (2008).

Diagnosis. HEINRICH (1926) stated that *Gymnandrosoma* is closely related to *Ecdytolopha* except for "spining of cucullus encroaching on neck of harpe; sacculus more weakly spined than neck".

In their revision of the genus, ADAMSKI & BROWN (2001) compared *Gymnandrosoma* to *Pseudogalleria* and *Cryptophlebia*. These authors and GILLIGAN et al. (2008) mentioned that the male scent scales are present on the abdomen terga, hind tibia and/or anal margin of the hindwing.

Remarks. Also mentioned under *Archiphlebia*, *Ecdytolopha*, *Lusterola*, *Pseudogalleria*, *Tachirinia*, and *Thaumatotibia*.

***Hyposarotis* DIAKONOFF, 1988**

Hyposarotis DIAKONOFF, 1988, Anns Soc. Ent. Fr. (N.S.), **24**(2): 168; t. sp.: *Hyposarotis atyphopa* DIAKONOFF, 1988, Madagascar. Two species included. **AFR**.

Redescription. RAZOWSKI (2004).

Diagnosis. DIAKONOFF (1988a) stated that *Hyposarotis* is “probably related to *Grapholita*, but distinct by the peculiar long brushes of hairs on the base of the hind wing dorsum and also by the entirely different facies: coloring and markings”.

***Ipamerica* RAZOWSKI & BECKER, 2016**

Ipamerica RAZOWSKI & BECKER, 2016, Zootaxa, **4066**(3): 253; t. sp.: *Ipamerica auctuncus* RAZOWSKI & BECKER, 2016, Brazil: Goias. One species included. **NEO**.

Diagnosis. In the original description *Ipamerica* is compared to *Ricula* from which it differs in having a distinct incision of the forewing termen beneath apex (lacking in *Ricula*) and in the reduction of the terminal row of spots which are conspicuous in *Ricula*.

***Ixonympha* KOMAI & HORAK, 2006**

Ixonympha KOMAI & HORAK, 2006, Monogr. Aust. Lepid., **10**: 464; t. sp.: *Hyphantidium hyposcopa* LOWER, 1905, Australia: Victoria. One species included. **AU**.

Diagnosis. According to KOMAI & HORAK (2006), *Ixonympha* is closely related to *Andrioplecta* and *Strophedra* “with the three genera sharing the following synapomorphies: (1) Sc+R1 and Rs entirely fused in the male; (2) two frenulum bristles in female; (3) M3 and CuA1 stalked in hindwing.... *Andrioplecta* is possibly the sister group of *Ixonympha*, sharing a bulla seminalis broadly connected to or continuous with the corpus bursae”. According to KOMAI (1999) “a pair of digital processes or flaps is present in the lateral tegumen wall of *Ixonympha* and several species of *Strophedra* and *Andrioplecta*”.

***Karacaoglania* KOÇAK, 1981**

Karacaoglania KOÇAK, 1981, Priamus, **1**: 115 – replacement name for *Diacantha* DIAKONOFF, 1976. One species included. **OR**.

Diacantha DIAKONOFF, 1976, Zool. Verh. Leiden, **144**: 42; t. sp.: *Laspeyresia xerophila* MEYRICK, 1939, India: Bihar, Pusa.

Diagnosis. According to DIAKONOFF (1976) *Diacantha* is “nearest to *Laspeyresia* (= *Cydia*)”.

Remarks. Male and female genitalia are accurately described and illustrated by DIAKONOFF in the original description.

***Larisa* MILLER, 1978**

Larisa MILLER, 1978, J. Lepid. Soc., **12**: 256; t. sp.: *Larisa subsolana* MILLER, 1978, USA: Arkansas. One species included. **NEA**.

Redescriptions. MILLER (1987), GILLIGAN et al. (2008)

Diagnosis. In the original description MILLER (1987) wrote: “Within *Laspeyresinae* *Larisa* most resembles *Lapeyresia* and *Hemimene* or *Pammene* (HEINRICH, 1926, OBRAZTSOV, 1960) but differs from both by its convex forewing termen, long setae on outer surface of cucullus, setal tufts of sacculus, well developed hamuli, and in previously enumerated details of forewings or hindwings neuration”.

***Lathronympha* MEYRICK, 1926**

Lathronympha MEYRICK, 1926, Entomologist, **59**: 27; t. sp.: [*Tortrix*] *hypericana* HÜBNER, [1799], Europe = *Pyralis strigana* FABRICIUS, 1775, Sweden. Seven species included. **PAL, AFR**.

Redescriptions. OBRAZTSOV (1960), DANILEVSKY & KUZNETZOV (1968), RAZOWSKI (1989, 2003).

Diagnosis. There is no original comparative diagnosis. DANILEVSKY & KUZNETZOV (1968) stated that *Lathronympha* is related to *Laspeyresia* (= *Cydia*), but has a separate position.

Based on wing venation, OBRAZTSOV (1960) suggested that *Lathronympha* is related to *Corticivora* but differs strongly in genitalia.

Remarks. Also mentioned under *Cydia*, *Coniostola* and *Leguminivora*.

***Leguminivora* OBRAZTSOV, 1960**

Leguminivora OBRAZTSOV, 1960, Tijdschr. Ent., **103**: 129; t. sp.: *Grapholitha glycinivorella* MATSUMURA, 1900, Japan: Hokkaido, Sapporo. Five species included. **PAL, OR, AFR, AU**.

Redescriptions. DANILEVSKY & KUZNETZOV (1968), RAZOWSKI (1989), KOMAI & HORAK (2006), NEDOSHIVINA (2013).

Diagnosis. OBRAZTSOV (1960) stated that *Leguminivora* differs from *Lathronympha* in the venation (forewing vein Cu₂).

DANILEVSKY & KUZNETZOV (1968) concluded that *Leguminivora* is related to *Cydia* and *Fulcrifera*.

KOMAI & HORAK (2006) compared *Leguminivora* to *Fulcrifera* (female sternal apodemes of S2 are stout, sternal rods in male at least vestigial), socii with long hairs (shared with *Notocydia*), arms of gnathos extending from below top of tegumen (as in *Fulcrifera*). Posterior parts of ovipositor lobes slender (as in *Notocydia*, *Fulcrifera*, and *Apocydia*).

Remarks. Also mentioned under *Acanthoclita*, *Amabrana*, *Cydia*, *Fulcrifera*, *Matsumuraes*, and *Notocydia*.

***Licigena* DIAKONOFF, 1982**

Licigena DIAKONOFF, 1982, Zool. Verh. Leiden, **193**: 13; t. sp.: *Licigena sertula* DIAKONOFF, 1982, Sri Lanka. One species included. **OR**.

D i a g n o s i s. There is no comparative original diagnosis. According to DIAKONOFF (1982) “the genus is characterised by relatively long, not dilated, curved and ascending labial palpi, and the rounded fore wing, while in the hind wing the veins 6 and 7 are separate and the veins 7 and 8 apparently coincident along basal half or more, so that vein 8 looks as a branch of 7”.

***Loranthacydia* HORAK, COMMON & KOMAI, 1996**

Loranthacydia HORAK, COMMON & KOMAI, 1996, Monogr. Aust. Lepid., **4**: 136; t. sp.: *Leptarthra aulacodes* LOWER, 1902, hereditarius. Replacement name for *Leptarthra* LOWER, 1902. Five species included. **AU**.

Leptarthra LOWER, 1902, Trans. R. Soc. S. Aust., **26**: 253; t. sp.: *Leptarthra aulacodes* LOWER, 1902, Australia: Western Australia. Junior homonym of *Leptarthra* BALY, 1861, Coleoptera.

D i a g n o s i s. KOMAI & HORAK (2006) provided an extensive diagnosis of *Loranthacydia* without a comparison to other genera.

LOWER (1902) diagnosed *Leptarthra* as follows: “Somewhat allied to *Byrsoptera*, LOWER, [= *Lobesia*, *Olethreutini*] but differing by the smooth thorax absence of secondary cell, and costa of hindwing”.

R e m a r k s. Also mentioned under *Commoneria*.

***Lusterola* BROWN & NISHIDA, 2007**

Lusterola BROWN & NISHIDA, 2007, Proc. Entomol. Soc. Wah., **109**(2): 266; t. sp.: *Lusterola phaseolana* BROWN & NISHIDA, 2007, Costa Rica. One species included. **NEO**.

D i a g n o s i s. BROWN & NISHIDA (2007) wrote that *Lusterola* is superficially most similar to *Gymnandrosoma*, *Ecdytolopha*, and *Thaumatotibia* by a dark brown forewing with few distinct pattern elements. The males lack secondary sexual scales typical of the above mentioned group of genera. These authors also compared the larvae of *Lusterola* and their biology.

***Macrocydia* BROWN & BAIXERAS, 2006**

Macrocydia BROWN & BAIXERAS, 2006, Zootaxa, **1197**: 46; t. sp.: *Macrocydia divergens* BROWN & BAIXERAS, 2006, Costa Rica. One species included. **NEO**.

D i a g n o s i s. BROWN & BAIXERAS (2006) wrote that *Macrocydia* is distinguished from all other Grapholitini by its conspicuously large size, its wing venation and colouration, and the absence of secondary sexual characters; they state that its

male genitalia are similar to many species of *Cydia* and *Grapholita*.

***Matsumuraeses* ISSIKI, 1957**

Matsumuraeses ISSIKI, 1957, Icones Heterocerorum Jap. Color. natural., **1**: 57; t. sp.: *Semasia phaseoli* MATSUMURA, 1900, Japan. Sixteen species included. **PAL, OR**.

R e d e s c r i p t i o n s. OBRAZTSOV (1960), DANILEVSKY & KUZNETZOV (1968), RAZOWSKI & YASUDA (1975, revision), RAZOWSKI (1989), KOMAI (1999, synopsis of the species), NEDOSHIVINA (2013).

D i a g n o s i s. Based on coremata OBRAZTSOV (1960) suggested that *Matsumuraeses* is related to *Grapholita*, and that *Pseudophiaris* and *Eucosmomorpha* (both belonging to Enarmoniini) are less specialized. OBRAZTSOV (1960) placed *Matsumuraeses* between *Leguminivora* and *Collicularia*, and DANILEVSKY & KUZNETZOV (1968) placed it at the beginning of their system. DIAKONOFF (1972) suggested that *Matsumuraeses* is closely related to *Cryptophlebia* and is placed at the end of his system.

In his diagnosis KOMAI (1999) compared *Matsumuraeses* to *Grapholita* (similar venation, “the valva is pincer-shaped in dorsal view, and the ringed or plate-shaped sclerite of the left side of the posterior end of corpus bursae”).

R e m a r k s. According to KOMAI (1999) “the monophyly of *Matsumuraeses* is supported by (1) the chorda from between R1 and R2 to between R5 and M1, (2) a pair of tufts of filiform scales on T7 in the male which is directed caudally and inserted into a pair of pouches on T8, and (3) the pincers-shaped valva”. His diagnosis does not contain a comparison to other genera.

Also mentioned under *Acanthoclita*, *Grapholita*, and *Pammenopsis*.

***Metacydia* RAZOWSKI & BECKER, 2012**

Metacydia RAZOWSKI & BECKER, 2012, Polish J. Entomol., **81**(3): 200; t. sp.: *Metacydia polyseta* RAZOWSKI & BECKER, 2012, Brazil: Rondonia. One species included. **NEO**.

D i a g n o s i s. In the original comparative diagnosis RAZOWSKI & BECKER (2012) compared the facies of *Metacydia* to those of *Ethelgoda*, *Ofatulena* and some *Cydia* from which it can be distinguished by the shape and vestiture of the valva. The numerous spiniform scales on the vinculum represent a male scent organ.

***Microsarotis* DIAKONOFF, 1982**

Microsarotis DIAKONOFF, 1982, Zool. Verh. Leiden, **193**: 10; t. sp.: *Laspeyresia palamedes* MEYRICK, 1916, India: Coimbatore. Seven species included. **OR, AFR, AU**.

Redescription. KOMAI & HORAK (2006).

Diagnosis. DIAKONOFF (1982) stated that *Microsarotis* is “an interesting genus with the venation of the hind wing unexpectedly resembling that in *Dichrorampha* GUENÉE, but with all external features of *Grapholita* TREITSCHKE, with coremata upon the seventh segment of the male but with quite distinct genitalia, viz., a distinct uncus and a peculiar broad and short valva”.

KOMAI (1999) and KOMAI & HORAK (2006) suggested that *Microsarotis* is a member of the *Grapholita*-group of genera.

Remarks. KOMAI (1999) included *Microsarotis* in the *Grapholita*-group of genera. According to KOMAI & HORAK (2006) the monophyly of *Microsarotis* is supported by several apomorphies including the wing venation, colouration, scent organs (the invaginated sacs with long scales angled proximally, etc.

Also mentioned under *Commoneria* and *Pammenitis*.

***Multiquestia* KARISCH, 2005**

Multiquestia KARISCH, 2005, Lambillionea, **105**: 500; t. sp.: *Multiquestia albimaculana* KARISCH, 2005, Angola. Nine species included. **AFR**.

Redescription. AARVIK & KARISCH (2009).

Diagnosis. AARVIK & KARISCH (2009) compared *Multiquestia* to *Cydia* and found two autapomorphies: the sclerotized anterior edge of female sternite 7 and “the hair pencil from the hind margin of the hind wing fitting under a scale ”roof” raised scales on the dorsal side of the abdomen in males.”

***Notocydia* KOMAI & HORAK, 2006**

Notocydia KOMAI & HORAK, 2006, Monogr. Aust. Lepid., **10**: 411; t. sp.: *Eucosma atripunctis* TURNER, 1946, Australia: Queensland. Four species included. **AU**.

Diagnosis. KOMAI & HORAK (2006) considered *Notocydia* to be the sister group of *Leguminivora*. These two genera share similar socii that have concavities from which arise long hair-like scales.

Remarks. Also mentioned under *Cydia* and *Leguminivora*.

***Ofatulena* HEINRICH, 1926**

Ofatulena HEINRICH, 1926, Bull. U. S. Natn. Mus., **132**: 41; t. sp.: *Grapholitha? duodecemstriata* WALSINGHAM, 1884, USA: Arizona. Eight species included. **NEA, NEO**.

Redescription. RAZOWSKI & BECKER (2012).

Diagnosis. HEINRICH (1926) originally described *Ofatulena* as “a small North American genus affiliated with *Laspeyresia* (= *Cydia*)”.

Remarks. Also mentioned under *Metacydia* and *Ranapoaca*.

***Pammene* HÜBNER, [1825]**

Pammene HÜBNER, [1825], Verz. bekannter Schmett.: 328; t. sp.: *Tortrix trauniana* [DENIS & SCHIFFERMÜLLER], 1775, Austria. Ninety species included. **PAL, OR, NEA**.

Palla BILLBERG, 1820, Enum. Insect.: 90; t. sp.: *Phalaena rhediella* CLERCK, 1759, Europe – preoccupied name.

Hemimene HÜBNER, [1825] 1816, Verz. bekannter Schmett.: 378; t. sp.: *Tortrix ephippiana* HÜBNER, [1817], Europe (= *Pyralis populana* FABRICIUS, 1787, Sweden).

Pseudotomia STEPHENS, 1829, Syst. Cat. Br. Insects, **2**: 175; t. sp.: *Phalaena (Tortrix) strobilella*: STEPHENS, 1829 [not LINNAEUS 1758] = *Tortrix argyrana* HÜBNER, [1796-1799], Europe.

Eucelis HÜBNER, [1825] 1816, Verz. bekannter Schmett.: 394; t. sp.: *Tortrix mediana* [DENIS & SCHIFFERMÜLLER], 1775, Austria (= *Pyralis aurana* FABRICIUS, 1775, Great Britain).

Heusimene STEPHENS, 1834, Illustr. Br. Ent. (Haustellata), **4**: 96; t. sp.: *Tortrix fimbriana* HAWORTH, [1811], Great Britain, preoccupied = *Coccyx giganteana* PEYERIMHOFF, 1863, France.

Encelis STEPHENS, 1834, Illustr. Br. Ent. (Haustellata), **4**: 105 – misspelling of *Eucelis*.

Pyrodes GUENÉE, 1845, Annl. Soc. Ent. Fr., (2)**3**: 187; t. sp.: *Phalaena rhediella* CLERCK, 1759, Europe.

Trycheris GUENÉE, 1845, Annl. Soc. Ent. Fr., (2)**3**: 190; t. sp.:

Tortrix mediana [DENIS & SCHIFFERMÜLLER], 1775, Austria (= *Pyralis aurana* Fabricius, 1775, Great Britain).

Orchemia GUENÉE, 1845, Annl. Soc. Ent. Fr., (2)**3**: 192; t. sp.: *Orchemia gallicana* GUENÉE, 1845, France.

Halonota STEPHENS, 1851, List Specimens Br. Anim. Colln Br. Mus., **10**: 45; t. sp. *Pyralis populana* FABRICIUS, 1787, Sweden.

Hemerosia STEPHENS, 1851, List Specimens Br. Anim. Colln Br. Mus., **10**: 60; t. sp.: *Phalaena rhediella* CLERCK, 1759, Europe – replacement name for *Palla* and *Pyrodes*.

Phthoroblastis LEDERER, 1859, Wien. Ent. Monatschr., **3**: 370; t. sp.: *Pyralis populana* FABRICIUS, 1787, Sweden.

Pamene REBEL, 1901, [in] STAUDINGER & REBEL Cat. Lepid. Pal. Faun., **2**: 123 – misspelling of *Pammene*.

Spharoea MEYRICK, 1895, Handbook Br. Lepid.: 490; t. sp.: *Pseudotomia obscurana* STEPHENS, 1834 – preoccupied.

Metasphaeroeca FERNALD, 1908, Gerena Tortricidae Types: 62 – replacement name for *Sphaeroeca*.

Eucells CARADJA, 1916, Dt. Ent. Z. Iris, **30**: 86 – misspelling of *Eucelis*.

Pammene MEYRICK, 1928, Exotic Microlepid., **3**: 447 – misspelling of *Pammene*.

Hemene PIERCE & METCALFE, 1935, Genitalia Tineid Families Lepid. Br. Is.: 114 – misspelling of *Hemimene*.

Redescriptions. KENNEL (1921), HEINRICH (1926 as *Hemimene*), OBRAZTSOV (1960), DANILEVSKY & KUZNETZOV (1968), RAZOWSKI (1989, 2003), KOMAI (1999), MILLER (1987), NEDOSHIVINA (2013).

Diagnosis. KENNEL (1921) compared *Pammene* to *Laspeyresia* (= *Cydia*).

HEINRICH (1926) compared *Hemimene* (= *Pammene*) to *Laspeyresia* (= *Cydia*): “Derived from and a higher development...”.

OBRATSOV (1960) stated that *Pammene* is very closely related to *Laspeyresia* (= *Cydia*) and that *Strophedra* differs from *Pammene* also in the genitalia.

DANILEVSKY & KUZNETZOV (1968) mentioned that *Pammene*, *Pammenodes* and *Parapammene* have similar venation in males.

Based on the venation KOMAI (1999) compared *Pammene* to *Pseudopammene*, *Dierlia*, and *Parapammene*.

Remarks. *Pammene* is usually divided into two subgenera, *Eucelis* and *Pammene* s. str., e.g. by DANILEVSKY & KUZNETZOV (1968) and RAZOWSKI (1989). The former authors distinguished *Eucelis* from *Pammene* s. str. by the position of scent scales on the male abdomen: segments 6-7 in *Eucelis*, segments 4-5 in *Pammene*.

KOMAI (1999) differentiated and characterized ten groups of species. He supposed that *Eucelis* is monophyletic but *Pammene* s. str. is paraphyletic. According to KOMAI the monophyly of *Pammene* is supported by: “(1) the sterigma formed by a rectangular plate with raised rim, (2) the short ductus bursae anteriorly with a cone-shaped ring, (3) the seventh sternite with a pair of triangular or round concavities laterally, and (4) T6 and T7 of males with a transverse patch of modified scales”.

Also mentioned under *Dierlia*, *Grapholita*, *Larisa*, *Pammenitis*, *Parapammene* and *Strophedra*.

***Pammenemima* DIAKONOFF, 1982**

Pammenemima DIAKONOFF, 1982, Zool. Verh. Leiden, **193**: 23; t. sp.: *Lipoptycha ochropa* MEYRICK, 1905, Sri Lanka. Five species included. **PAL, OR, AFR, AU.**

Titanotoca DIAKONOFF, 1984, Entomol. Basil., **9**: 380; t. sp.: *Titanotoca pagerostoma* DIAKONOFF, 1984, Sumba.

Redescription. HORAK & KOMAI (2006).

Diagnosis. *Pammenemima* was compared by DIAKONOFF (1982, described in Eucosmini) to *Pammenodes* on the basis of venation of the female, “differing from that genus by the unusual vein inside the cell of the hind wing... Peculiar are the female genitalia, characterised by the presence of a kind of a second, miniature, bursa copulatrix, opening immediately beside the ostium bursae”.

KOMAI & HORAK (2006) pointed that *Pammenemima* shares with *Dichrorampha* three or four black terminal spots, the hindwing venation with Rs and M1 parallel and distant at base, the

short apical process of the tegumen, and the sterigma fused with S7.

In the original description DIAKONOFF (1982) characterized *Titanotoca* as follows: “a small insect with a general facies of a *Cydia*, except for unusual, curved and rising, long palpi and absence of a cubital pecten. The genitalia are peculiar because of the very large and sclerotic, scobinate aedeagus”.

***Pammenitis* DIAKONOFF, 1988**

Pammenitis DIAKONOFF, 1988, Anns Soc. Ent. Fr. (N.S.), **24**(2): 167; t. sp.: *Pammenitis calligrapha* DIAKONOFF, 1988, Madagascar. One species included. **AFR.**

Redescription. RAZOWSKI (2004).

Diagnosis. According to DIAKONOFF (1988a) *Pammenitis* is a close relative of *Pammenodes*, “but differing in the presence of slight coremata and absence of abdominal androconia while the peculiar armature of the valva forms a secondary autapomorphy: the huge submarginal spines at the base of the sacculus and the dense spiny armour of the broad edge of sacculus and cucullus. From *Pammene* HÜBNER, both these genera differ by the neuration of the hind wing being similar in the two sexes”.

According to RAZOWSKI (2004) *Pammenitis* resembles *Microsarotis* but has a different cucullar part of the valva, armoured with marginal thorns.

***Pammenopsis* KUZNETZOV, 2003**

Pammenopsis KUZNETZOV, 2003, Entomol. Obozr., **82**: 740; t. sp.: *Eucelis critica* MEYRICK, 1905, India: Bombay. Two species included. **OR, AU.**

Redescriptions. KOMAI & HORAK (2006), NEDOSHIVINA (2013).

Diagnosis. KUZNETZOV (2003) – compared *Pammenopsis* to *Pammene*; the two have similar but differently arranged androconial structures and valvae. Based on the genitalia *Pammenopsis* is similar to *Matsumuraeses* but has a different cucullus.

According to KOMAI & HORAK (2006) *Pammenopsis* is similar to *Pammene* and *Matsumuraeses* but the former is distinct superficially, in the venation and genitalia (the autapomorphies are listed).

***Parapammene* OBRATSOV, 1960**

Parapammene OBRATSOV, 1960, Tijdschr. Entomol., **103**: 125; *Phthoroblastis selectana* CHRISTOPH, 1882, E Russia: Amur District. Seventeen species included. **PAL, OR, AU.**

Diamphidia OBRATSOV, 1961, Tijdschr. Entomol., **104**: 51; t. sp.: *Pammene petulantana* KENNEL, 1901, Russia: Amur Region.

Pammenodes DANILEVSKY & KUZNETZOV, 1968, Fauna SSSR (N.S.), **98**, (Lepid.,5(1)): 334; t. sp.: *Pammene glaucana* KENNEL, 1901, Russia: Primorsky Krai.

Mimarsinania KOÇAK, 1981, Priamus, **1**: 116. Replacement name for *Diamphidia* OBRAZTSOV, 1961.

Diplosemaphora DIAKONOFF, 1982, Zool. Verh. Leiden, **193**: 35; t. sp.: *Diplosemaphora amphibola* DIAKONOFF, 1982, Sri Lanka.

Redescriptions. DANILEVSKY & KUZNETZOV (1968, also as *Diamphidia*), RAZOWSKI (1989), KOMAI (1999), KOMAI & HORAK (2006).

Diagnosis. OBRAZTSOV (1960) described *Parapammene* as externally similar to *Pammene* from which it differs in the scent organs; it differs from *Strophedra* by the possession of an appendix bursae in the female genitalia.

DANILEVSKY & KUZNETZOV (1968) mentioned that the androconia of *Parapammene* and *Diamphidia* resemble those of *Grapholita* but the venation is different, similar to that of *Strophedra*.

According to DANILEVSKY & KUZNETZOV (1968) *Pammenodes* is related to *Parapammene* sharing facies, tegumen and eighth abdominal tergite of males but differing from it by a lack of coremata, ventral androconial organ (of a *Pammene* type) and fully developed venation of the hindwing. Females of *Pammenodes* differ from those of *Parapammene* by a lack of a parabursa and sclerite of ductus bursae.

KOMAI (1999) compared the venation of *Parapammene* to that of *Pseudopammene*, *Dierlia* and *Strophedra*. He listed the autapomorphies of *Parapammene* (see below) and mentioned that they are shared by *Parapammene* and *Pammenodes*. KOMAI observed that “the male hindwing venation of *Mimarsinania* and both genera [*Pammenodes* and *Parapammene*] are derivations of the basic plan of *Pammene* + *Dierlia* + *Pseudopammene* + *Parapammene* + *Strophedra* + *Andrioplecta* which is also shared by the other members of *Parapammene* as defined here”.

Remarks. KOMAI (1999) mentioned the following putative autapomorphies which support the monophyly of *Parapammene*: (1) the ductus bursae with a narrow, band-like sclerite, (2) seventh sternite of female a convex (not flat) plate, and (3) S2 without anterolateral process”.

Also mentioned under *Andrioplecta*, *Grapholita*, *Dierlia*, *Diamphidia*, *Pammenodes* and *Strophedra*.

***Phloerampha* RAZOWSKI, 2011**

Phloerampha RAZOWSKI, 2011, Acta zool. cracov., **53**(1-2): 48; t. sp.: *Phloerampha phloea* RAZOWSKI, 2011, Venezuela. Three species included. **NEO**.

Diagnosis. In the original description RAZOWSKI (2011a), *Phloerampha* was placed in

the *Dichrorampha*-group; the male genitalia were compared to those of *Cydia* and the female genitalia to *Sereda* but the sterigma of the latter is broad and short; compared to *Ethelgoda* the sterigma of *Phloerampha* differs in having distal, concave folds whilst in *Ethelgoda* these parts are convex.

***Pseudogalleria* RAGONOT, 1884**

Pseudogalleria RAGONOT, 1884, Anns Soc. Ent. Fr., (6)4 (Bulletin): L; t. sp.: *Galleria inimicella* ZELLER, 1872, USA: Texas. One species included. **NEA**.

Redescriptions. HEINRICH (1926), MILLER (1987), ADAMSKI & BROWN (2001), GILLIGAN et al. (2008).

Diagnosis. GILLIGAN et al. (2008) compared *Pseudogalleria* to *Gymnandrosoma* and *Ecdytolopha*; the tegumens are very similar but the “sacculus lacks stout setae on its distal margin but has several setae that are distributed along a distinctive ridge along medial surface...”. In the phylogenetic tree, ADAMSKI & BROWN (2001) concluded that *Pseudogalleria* is the sister genus of *Cryptophlebia*.

Based on facies and male genitalia, it is highly likely that *Pseudogalleria* is synonymous with *Cryptophlebia*, as suggested by HORAK & KOMAI (2016), and *Pseudogalleria* is the senior synonym.

***Pseudopammene* KOMAI, 1980**

Pseudopammene KOMAI, 1980, Tinea, **11**(1): 2; t. sp.: *Pseudopammene fagivora* KOMAI, 1980, Japan: Honsyu. One species included. **PAL**.

Redescriptions. KOMAI (1999), RAZOWSKI (1989).

Diagnosis. KOMAI (1980) regarded *Pseudopammene* as the sister group of *Dierlia*; *Pseudopammene* and *Dierlia* also are related to *Agriophanes*.

Remarks. Also mentioned under *Agriophanes*, *Dierlia*, and *Parapammene*.

***Ranapoaca* RAZOWSKI, 2011**

Ranapoaca RAZOWSKI, 2011, Acta zool. cracov., **53A**(1-2): 56; t. sp.: *Ranapoaca caparoana* RAZOWSKI, 2011, Mexico. Four species included. **NEO**.

Diagnosis. *Ranapoaca* is most closely related to *Ricula* on the basis of shared fusion of the sternum with the sterigma and the possession of one signum in the corpus bursae (the latter shared also with *Goditha*, *Riculorampha* and many *Dichrorampha*) and a broad aedeagus. *Ranapoaca* is similar to some species of *Ofatulena*.

***Ricula* HEINRICH, 1926**

Ricula HEINRICH, 1926, Bull. U.S. Natn. Mus., **132**: 18; t. sp.: *Lipoptycha maculana* FERNALD, 1901, USA: Florida. Over 40 species included. **NEO**.

Riculoides PASTRANA, 1952, Bull. Soc. Cient. Argent., **154**: 66; t. sp.: *Riculoides gallicola* PASTRANA, 1952, Argentina.

Redescriptions. RAZOWSKI (2011a), RAZOWSKI & BECKER (2011).

Diagnosis. HEINRICH (1926) diagnosed *Ricula* as follows: “related to *Talponia*, from which it differs chiefly in the more approximate condition of veins 6 and 7 of hindwing, the convexity of the termen of fore wing, and the presence of one signum from the bursa of the female”.

PASTRANA (1952) compared *Riculoides* to *Ricula* (differing in shape of hindwing, venation, longer socii in the former, and ductus bursae).

Remarks. RAZOWSKI (2011a) included *Ricula* in the *Dichrorampha*-group of genera which have only one signum. Males of this genus have a pair of strong setae at the top of the tegumen and long, slender socii.

Also mentioned under *Acailandica*, *Balbis*, *Ethelgoda*, *Ipamerica*, and *Ranapoaca*.

***Riculorampha* ROTA & BROWN, 2009**

Riculorampha ROTA & BROWN, 2009, ZooKeys, **23**: 41; t. sp.: *Riculorampha ancyloides* ROTA & BROWN, 2009, USA: Florida. Three species included. **NEA**.

Redescription. RAZOWSKI (2011a).

Diagnosis. In the original description ROTA & BROWN 2009 *Riculorampha* was compared to *Dichrorampha*, *Goditha*, *Ricula* and *Riculoides* which possess only one signum. In the male genitalia this genus differs from *Ricula* by the reduction of socii, rounded dorsum of the tegumen and large, triangular sacculus which is found also in *Goditha* and *Dichrorampha*.

***Satronia* HEINRICH, 1926**

Satronia HEINRICH, 1926, Bull. U. S. Natn. Mus., **132**: 17; t. sp.: *Satronia tantilla* HEINRICH, 1926, USA: Florida. Twelve species included. **NEO**.

Redescription. RAZOWSKI (2011a).

Diagnosis. In the original description (HEINRICH, 1926) *Satronia* is regarded as “a higher development from *Ricula*. The male genitalia are similar in both except for the socii”.

RAZOWSKI (2011a) compared the female genitalia of *Satronia* to those of *Talponia* and *Sereda* which share the presence of two signa in the corpus bursae. The male genitalia of *Satronia* are most similar to those of *Talponia*, but *Satronia* lacks setae at the top of the tegumen and long, slender socii.

Remarks. Also mentioned under *Acailandica*, *Ricula*, *Sereda*, and *Talponia*.

***Selania* STEPHENS, 1834**

Selania STEPHENS, 1834, Illust. Br. Ent. Haustellata, **4**: 121; t. sp.: *Carpocapsa leplastriana* CURTIS, 1831, Great Britain. Fourteen species included. **PAL, AFR, OR**.

Chretienia OBRAZTSOV, 1968, J. New York Ent. Soc., **76**: 224; t. sp.: *Grapholitha rhezelana* CHRÉTIEN, 1915, Algeria = *Grapholitha capparidana* ZELLER, 1847, Italy, a junior homonym of *Cretienia* SPULER, 1910, Lepidoptera, Gelechiidae.

Mevlanaia KOÇAK, 1981, Priamus, **1**: 115, replacement name for *Chretienia*.

Redescriptions. DANILEVSKY & KUZNETZOV (1968), DIAKONOFF (1984, *Chretienia*), RAZOWSKI (1989, 2003, 2004), KOMAI (1999).

Diagnosis. According to DANILEVSKY & KUZNETZOV (1968) the genitalia of *Selania* are similar to those of *Grapholita*.

Remarks. KOMAI (1999) stated that the monophyly of *Selania* is supported by “(1) the valva being pincers-shaped in dorsal view, (2) the ductus bursae with a sclerotized ring in the anterior end, (3) the corpus bursae with an elongate, ridged or plated sclerite on the left side of the posterior end (...sometimes atrophied), and (5) the reduced signum (usually it [is] absent)”. Two groups of species were proposed, the *leplastriana*-group and the *capparidana*-group.

Remarks. Also mentioned under *Grapholita*.

***Sereda* HEINRICH, 1923**

Sereda HEINRICH, 1923, Proc. Ent. Soc. Washington, **25**: 121; t. sp.: *Halonota lautana* [as *tautana*] CLEMENS, 1865, USA: Virginia. Twelve species included. **NEO**.

Redescriptions. HEINRICH (1926), MILLER (1987), GILLIGAN et al. (2008).

Diagnosis. According to HEINRICH (1926) *Sereda* is “a monotypic genus derived from *Grapholita* (= *Grapholita*). The absence of the pecten is rare for the family, occurring elsewhere, as far as I know, only in *Satronia* and *Goditha*”.

Remarks. Also mentioned under *Phloerampha*, *Satronia*, and *Talponia*.

***Spanistoneura* DIAKONOFF, 1982**

Spanistoneura DIAKONOFF, 1982, Zool. Verh. Leiden, **193**: 8; t. sp.: *Spanistoneura acrospecta* DIAKONOFF, 1982, Sri Lanka. One species included. **OR**.

Spartoneura DIAKONOFF, 1982, Zool. Verh. Leiden, **193**: 9 – misspelling of *Spanistoneura*.

Diagnosis. According to DIAKONOFF (1982) *Spanistoneura* is “an interesting form with peculiarly specialised female genitalia and reduced wing neuration, obviously belonging to the *Gra-*

pholita TREITSCHKE group of the subtribe Cydiae, but considerably modified”.

Stephanopyga DIAKONOFF, 1988

Stephanopyga DIAKONOFF, 1988, Anns Soc. Ent. Fr. (N. S.), **24**(2): 175; t. sp.: *Stephanopyga legnota* DIAKONOFF, 1988, Madagascar. One species included. **AFR**.

Redescription. RAZOWSKI (2004).

Diagnosis. DIAKONOFF (1988a) stated that *Stephanopyga* is “a peculiar form, as to the colouring resembling slightly *Grapholita miranda* (MEYRICK), but otherwise completely different. Unusual are the long, loosely haired palpi and the male genitalia”.

According to RAZOWSKI (2004) *Stephanopyga* resembles *Cirriphora*. The male genitalia have terminal, rather well developed socii and broad valva devoid of a neck and with a very small cucullus.

Strophedra HERRICH-SCHÄFFER, 1853

Strophedra HERRICH-SCHÄFFER, 1853, Syst. Bearbeitung Schmett. Eur., **5**: 94; t. sp.: *Strophedra vigeliana* HERRICH-SCHÄFFER, 1853, Europe: Germany = *Pyralis nitidana* FABRICIUS, 1974, Europe: Great Britain. Replacement name for *Strophosoma* HERRICH-SCHÄFFER, 1853. Nine species included. **PAL, OR, AU**.

Strophosoma HERRICH-SCHÄFFER, 1853, Syst. Bearbeitung Schmett. Eur., **5**: 8, 29; t. sp.: *Strophedra vigeliana* HERRICH-SCHÄFFER, 1853, Europe: = *Pyralis nitidana* Fabricius, 1974, Europe: Great Britain – preoccupied.

Strophedromorpha DIAKONOFF, 1976, Zool. Verh. Leiden, **144**: 29; t. sp.: *Strophedromorpha mica* DIAKONOFF, 1976, Nepal.

Redescriptions. DANILEVSKY & KUZNETZOV (1968), OBRAZTSOV (1960), RAZOWSKI (1989, 2003), KOMAI (1999).

Diagnosis. OBRAZTSOV (1960) compared the genitalia of *Strophedra* to those of *Pammene* indicating that they differ by the laterally scaled coremata of the 8th abdominal segment, and that females of *Strophedra* have a “ventral plate with small median process”.

DANILEVSKY & KUZNETZOV (1968) mentioned that the male genitalia of *Strophedra* resemble those of the *Grapholita*-group, with a similar fusion of veins R-M1 in male hindwings as in *Diamphidia*. According to DIAKONOFF (1976) *Strophedromorpha* “resembles a Palaearctic *Strophedra* closely, but the genitalia are very peculiar and the vein 7 in the hind wing is absent”.

RAZOWSKI (1989) compared the male genitalia of *Strophedra* to those of *Grapholita* and treated the presence of the process of the posterior edge of the subgenital sternite as an autapomorphy for this genus.

KOMAI (1992, 1999) noted that *Strophedra* and *Andrioplecta* “are closely related as indicated by

the female frenulum consisting of two bristles and the ductus bursae narrow and almost entirely sclerotized with a longitudinal groove”.

Remarks. KOMAI (1999) stated that the following characters support the monophyly of the genus: “T8 of male with a pair of tufts of long hair-like scales arising from shallow membranous pockets on each side and S7 of female with posterior edge produced into a median process”.

Based on larval characters SWATSCHKE (1958) included the species of *Strophedra* in *Pammene*.

Remarks. Also mentioned under *Andrioplecta*, *Ixonympha*, *Pammene* and *Parapammene*.

Tachirinia RAZOWSKI & WOJTUSIAK, 2013

Tachirinia RAZOWSKI & WOJTUSIAK, 2013, Acta zool. cracov., **56**(1): 28; t. sp.: *Tachirinia rosallana* RAZOWSKI & WOJTUSIAK, 2013, Venezuela: Paramo el Rosal. One species included. **NEO**.

Diagnosis. Originally, (RAZOWSKI & WOJTUSIAK, 2013) mentioned that *Tachirinia* is related to *Gymnandrosoma* but *Tachirinia* has well developed, slender socii and a transverse row of setae beyond the end of the valva.

Talponia HEINRICH, 1926

Talponia HEINRICH, 1926, Bull. U. S. Natn Mus., **132**: 19; t. sp.: *Hemimene plummeriana* BUSCK, 1906, USA: Maryland, Plummers Island. Nine species included. **NEA, NEO**.

Redescriptions. MILLER (1987), GILLIGAN et al. (2008), RAZOWSKI (2011a), RAZOWSKI & BECKER (2011).

Diagnosis. HEINRICH (1926) described *Talponia* as “allied to *Ricula*, *Ethelgoda*, and the tropical *Balbis* WALSINGHAM. In wing shape, general habitus most like *Ethelgoda*. In genitalia (male and female) closest to *Balbis*...”.

RAZOWSKI (2011a) compared *Talponia* to *Satronia* and *Sereda* which have two signa, and to *Riculoides* on the basis of the narrow, dorsally attenuate tegumen with setae at the top, and the long, slender socii. The genitalia of *Talponia* also are similar to those of *Ricula*, but in *Talponia* the tegumen is more attenuate dorsally, the socii are much longer, and the sterigma is fused with subgenital sternite.

Remarks. Also mentioned under *Ethelgoda*, *Ricula* and *Satronia*.

Thaumatotibia ZACHER, 1915

Thaumatotibia ZACHER, 1915, TROPENPFLANZER, **18**: 529; t. sp.: *Thaumatotibia roerigerii* ZACHER, 1915, Togo = *Argyroploce leucotreta* MEYRICK, 1913, South Africa: Transvaal, Pretoria. Twenty species included. **PAL, OR, AFR, AU**.

Metriophlebia DIAKONOFF, 1969, Tijdschr. Entomol., **112**(3): 89; t. sp.: *Eucosma chaomorpha* MEYRICK, 1929, Marquesas.

Redescriptions. KOMAI (1999), ADAMSKI & BROWN (2001, phylogeny), RAZOWSKI (2004), KOMAI & HORAK (2006), RAZOWSKI & BROWN (2012).

Diagnosis. According to KOMAI (1999) “*Thaumatotibia* has relationships with *Cryptophlebia*, *Ecdytolopha*, and *Pseudogalleria* as indicated by the common possession of the following autapomorphies: forewing [shape and colouration] and with accessory cell of chorda small or absent (chorda coincident with margin of discal cell); hindwing with a short discal cell especially in male; T8 and sometimes also preceding tergites bearing a patch of long mane-like scales; valva with a patch of very long, curled scales on outer surface of cucullus;” etc.

RAZOWSKI (2004) and KOMAI (1999) mentioned that *Thaumatotibia* is characterized chiefly by the sclerotized male subgenital tergite with convex distal edge and broad convexity of the terminal portion of corpus bursae.

KOMAI & HORAK (2006) compared *Thaumatotibia* to *Cryptophlebia*, *Pseudogalleria*, *Gymnandrosoma*, and *Ecdytolopha*.

DIAKONOFF (1969) described *Metriophlebia* as a genus closely allied with *Cryptophlebia* from which it differs in the facies, denticulate juxta and the venation (“peculiar origin of vein 2 and absence of chorda in the forewing and by the stalking of veins 3 and 4 in hind wing”).

Remarks. KOMAI (1999) provided a synopsis of the included species.

Also mentioned under *Archiphlebia*, *Celsumaria*, *Cryptophlebia*, *Dracontogena*, *Ecdytolopha*, *Lusterola*, and *Thaumatovalva*.

Thaumatovalva TIMM & BROWN, 2014

Thaumatovalva TIMM & BROWN, 2014, ZooKeys, **438**: 113-128; t. sp.: *Thaumatovalva albolineana* TIMM & BROWN, 2014, Democratic Republic of Congo: Rutshuru. Four species included. **AFR**.

Diagnosis. Originally (TIMM & BROWN, 2014), *Thaumatovalva* was compared to *Thaumatotibia* and *Cryptophlebia* from which it differs in the hindwing scent scales. The genital differences are of the lesser importance.

Thylacandra DIAKONOFF, 1963

Thylacandra DIAKONOFF, 1963, Verh. Naturf. Ges. Basel, **74**(1): 142; t. sp.: *Retinia argyromixtana* MABILILE, 1900, Madagascar. Five species included. **AFR**.

Thylacandra DIAKONOFF, 1963, Verh. Naturf. Ges. Basel, **74**(1): 142 – misspelling of *Thylacandra*.

Thylacandra DIAKONOFF, 1963, Verh. Naturf. Ges. Basel, **74**(1), pl. 3 – misspelling of *Thylacandra*.

Redescription. RAZOWSKI (2004).

Diagnosis. According to the original description *Thylacandra* was characterized as “allied with *Cryptophlebia* WALS”. Its valva has a peculiar, rounded organ situated near middle subcostally comparable to that of *Dracontogena*.

Based on facies and male genitalia, in particular the orbicular process in the middle of the valva, *Celsumaria* is most likely a synonym of *Thylacandra*.

Remarks. Also mentioned under *Celsumaria* and *Dracontogena*.

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INDEX TO SYNONYMS

- Adenoneura* - *Cydia*
Amaurosetia - *Dichrorampha*

Carpocampa - *Cydia*
Carpocapsa - *Cydia*
Cerata - *Cydia*
Cirriphora - *Coccothera*
Chretienia - *Selania*
Coccyx - *Cydia*
Collicularia - *Cydia*
Coptoloma - *Grapholita*, *Aspila*
Crobylophora - *Cydia*

Danilevskia - *Cydia*
Diacantha - *Karacaoglania*
Diamphidia - *Parapammene*
Dichroramphodes - *Dichrorampha*
Dicrorampha - *Dichrorampha*
Dicraniana - *Cydia*
Diplosemaphora - *Parapammene*

Ebisma - *Grapholita*
Ecdytolophia - *Ecdytolopha*
Encelis - *Pammene*
Endopisa - *Grapholita*
Endothera - *Coccothera*
Ephippiphora - *Grapholita*
Erminea - *Cydia*
Eucelis - *Pammene*
Euspila - *Grapholita*

Grapholitha - *Grapholita*
Gynandrosoma - *Gymnandrosoma*

Halonota - *Pammene*
Hemerosia - *Pammene*
Hedulia - *Cydia*
Hemene - *Pammene*
Hemimene - *Pammene*
Heusimene - *Pammene*

Kenneliola - *Cydia*

Laspeyresia - *Cydia*
Lepidoptycha - *Dichrorampha*

Leptarthra - *Loranthacydia*
Lipoptycha - *Dichrorampha*
Lipoptychodes - *Dichrorampha*

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Mellisopus - *Cydia*
Mellissopus - *Cydia*
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Mesotis - *Acanthoclita*
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